



The InBIO Barcoding Initiative Database: contribution to the knowledge on DNA barcodes of cuckoo wasps, with the description of new species from the Iberian Peninsula (Hymenoptera, Chrysididae)

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Abstract

Background

DNA barcoding technologies have provided a powerful tool for the fields of ecology and systematics. Here, we present a part of the InBIO Barcoding Initiative Database: contribution to the knowledge on DNA barcodes of cuckoo wasps (Hymenoptera, Chrysididae) dataset representing 144 specimens and 103 species, covering

approximately 44% of the Iberian and 21% of the European fauna. The InBIO Barcoding Initiative (IBI – DNA Barcoding Portuguese terrestrial invertebrate biodiversity) aims to fill the barcoding gap for the terrestrial invertebrate taxa. All DNA extractions are deposited in the IBI collection at CIBIO, Research Center in Biodiversity and Genetic Resources and specimens are deposited in the University of Mons collection (Belgium) and in the Natur-Museum in Lucerne (Switzerland).

New information

This dataset increases the knowledge on the DNA barcodes and distribution of 102 species of cuckoo wasps. A total of 52 species, from 11 different genera, were new additions to the Barcode of Life Data System (BOLD), with DNA barcodes for another 44 species added from under-represented taxa in BOLD. All specimens have their DNA barcodes publicly accessible through the BOLD online database. Nine cuckoo wasp species are newly recorded for Portugal. Additionally, two new species for science are described: *Chrysis crossi* Rosa, sp. nov. from southern Portugal and *Hedychridium calcarium* Rosa, sp. nov. from eastern Spain. Several taxonomic changes are proposed and *Hedychrum rutilans* Dahlbom, 1845 is found to consist of two different taxa that can be found in sympatry, *Hedychrum rutilans* s. str. and *Hedychrum viridaureum* Tournier, 1877 stat. nov. *Stilbum westermanni* Dahlbom, 1845 stat. nov. is confirmed as distinct from *Stilbum calens* (Fabricius, 1781), with the latter species not confirmed as present in Iberia; barcoded *Stilbum* material from Australia is distinct and represents *Stilbum amethystium* (Fabricius, 1775) sp. resurr.; Portuguese material identified as *Hedychridium chloropygum* Buysson, 1888 actually belongs to *Hedychridium caputaureum* Trautmann & Trautmann, 1919, the first confirmed record of this species from Iberia. *Philoctetes parvulus* (Dahlbom, 1845) is confirmed to be a synonym of *Philoctetes punctulatus* (Dahlbom, 1845). *Chrysis lusitanica* Bischoff, 1910 is confirmed as a valid species. *Chrysis hebraeica* Linsenmaier, 1959 stat. nov. is raised to species status.

Keywords

Portugal, Spain, Italy, DNA barcode, mitochondrial DNA, Cytochrome c oxidase subunit I (COI)

Introduction

In Europe, the diversity of cuckoo wasps is highest in the Mediterranean region, with relatively few species found in the north (Paukkunen et al. 2014, Paukkunen et al. 2015) and the British Isles (Morgan 1984). Chrysidids are more common in southern European countries in part due to their ecology, since most species are heliophilous and thermophilous, favouring warm and sunny habitats. Another reason is their reproductive biology, as the number of host species of bees and aculeate wasps is also greater in Mediterranean countries (Michez et al. 2019).

The total number of valid cuckoo wasp species is approximately 2,800 (Rosa 2017). Of this world total, about 480 have been recorded from Europe, plus 135 accepted subspecies, whose possible specific rank has yet to be evaluated (Mitroiu et al. 2015). In Portugal, a total of 130 species and four subspecies are known to date, but this number is likely to be far from the true total given the much larger number of species reported from neighbouring Spain (e.g. González et al. 1999, González et al. 2009, Mingo and Gayubo 1981, Mingo and Gayubo 1986). Mingo (1994) compiled the most recent faunistic survey for the Iberian Peninsula, including identification keys. In this important monograph, Portuguese species are not clearly separated from Spanish ones. This volume is a valuable guide for beginners, yet includes only 170 species for the Iberian Peninsula, overlooking many of the species described or published from Spain. The real number of species exceeds 230 known taxa. However, new field research on Portuguese bees and aculeate wasps (e.g. Rosa et al. 2015, Rosa et al. 2015, Rosa and Vårdal 2015, Rosa and Xu 2015, Baldock et al. 2018, Baldock et al. 2020, Cross et al. 2021) has reinvigorated work on the Iberian cuckoo wasp fauna and a new illustrated catalogue of the Portuguese fauna is in preparation, including new records for the country and for Europe.

Despite the fact that the Iberian Chrysididae fauna is one of the richest in Europe (Mingo 1994), this fauna has essentially never been investigated using molecular tools, with only a handful of DNA barcodes sequences available from a small selection of species (e.g. Pauli et al. (2019)). To a certain extent, this is a function of the limited number of studies that have presented barcode data for West Palaearctic Chrysididae in general (Niehuis and Wägele 2004, Soon and Saarma 2011, Soon et al. 2014, Paukkunen et al. 2015, Orlovskaitytė et al. 2016, Roslin et al. 2021). The present work represents the first attempt to generate DNA barcodes for Iberian Chrysididae at a faunal level and, thus, represents a major step in documenting the genetic diversity in the Mediterranean cuckoo wasp fauna.

Materials and methods

This dataset is composed of data relating to 144 Chrysididae specimens. Specimens were collected during field expeditions in the Iberian Peninsula, Belgium, Italy and Morocco from 2014 to 2022 by T.J. Wood, I. Cross (Dorchester, UK) and P. Rosa (Fig. 1, Table 1). Specimens were pinned and dried and are preserved in the collection of T.J. Wood at the University of Mons (Belgium); Italian specimens are preserved in ethanol (98%) and preserved in the collection of P. Rosa at the University of Mons (Belgium). Holotypes of the newly-described species are deposited in the Natur-Museum (Lucerne, Switzerland – **NMLU**) and paratypes are deposited in the following private collections: **PRC** (Paolo Rosa Collection, Bernareggio, Italy), **TWC** (Thomas J. Wood Collection, Mons, Belgium); **ICC** (Ian Cross Collection, Briantspuddle, Dorset, United Kingdom). The majority of specimens were determined to species level, though some specimens in challenging or unclear taxonomic groups were identified as 'cf.' or simply to the species group. Overall, 103 species are represented in the dataset. These species belong to 13 genera (Fig. 2).

Table 1.

List of species that were collected and DNA barcoded within this project. # Indicates species with new BINs.

| Genus | Species | IBI code | BOLD code | BOLD BIN | GenBank |
|------------------|--|----------|--------------|------------------------------|--------------------------|
| <i>Chrysidea</i> | <i>Chrysidea disclusa pumilionis</i> (Lisenmaier, 1987) # | INV12702 | IBIHM1103-22 | BOLD:AES3051 | OP347205 |
| <i>Chrysis</i> | <i>Chrysis andradei</i> Lisenmaier, 1959# | INV12728 | IBIHM1129-22 | BOLD:AES8383 | OP347228 |
| | | INV12729 | IBIHM1130-22 | | OP347302 |
| | <i>Chrysis berlandi</i> Lisenmaier, 1959# | INV12682 | IBIHM1083-22 | BOLD:AES5679 | OP347200 |
| | | INV12683 | IBIHM1084-22 | | OP347265 |
| | | INV12734 | IBIHM1135-22 | | OP347250 |
| | <i>Chrysis blanchardi</i> Lucas, 1849# | INV12732 | IBIHM1133-22 | BOLD:AEU3313 | OP347222 |
| | <i>Chrysis caeruliventris</i> Abeille de Perrin, 1878 | INV12676 | IBIHM1077-22 | BOLD:AED3523 | OP347173 |
| | <i>Chrysis castillana</i> Du Buysson, 1894 | INV12731 | IBIHM1132-22 | BOLD:AED2289 | OP347274 |
| | <i>Chrysis cerastes</i> Abeille de Perrin, 1877# | INV12733 | IBIHM1134-22 | BOLD:AET4960 | OP347293 |
| | <i>Chrysis rutilans</i> Olivier, 1790# | INV12689 | IBIHM1090-22 | BOLD:AET4959 | OP347268 |
| | <i>Chrysis chrysoprasina</i> Forster, 1853 | INV12677 | IBIHM1078-22 | BOLD:AET4958 | OP347283 |
| | <i>Chrysis chrysoscutella</i> Lisenmaier, 1959# | INV12743 | IBIHM1144-22 | BOLD:AES1459 | OP347305 |
| | <i>Chrysis comparata</i> Lepeletier, 1806 | INV12679 | IBIHM1080-22 | BOLD:AAU1528 | OP347241 |
| | <i>Chrysis consanguinea</i> Mocsáry, 1889 | INV12687 | IBIHM1088-22 | BOLD:AED0671 | OP347212 |
| | <i>Chrysis cortii</i> Lisenmaier, 1951 | INV12670 | IBIHM1071-22 | BOLD:AAR9816 | OP347211 |
| | <i>Chrysis elegans</i> Lepeletier, 1806# | INV12673 | IBIHM1074-22 | BOLD:AES1460 | OP347219 |
| | <i>Chrysis emarginatula</i> Spinola, 1808 | INV12672 | IBIHM1073-22 | BOLD:AED6786 | OP347309 |
| | <i>Chrysis fugax</i> Abeille de Perrin, 1878 | INV12661 | IBIHM1062-22 | BOLD:AED3372 | OP347230 |
| | <i>Chrysis germari</i> Wesmael, 1839# | INV12669 | IBIHM1070-22 | BOLD:AET6935 | OP347229 |
| | <i>Chrysis gracillima aurofacies</i> (Trautmann, 1926) # | INV12663 | IBIHM1064-22 | BOLD:AES2863 | OP347234 |
| | <i>Chrysis grohmanni</i> Dahlbom, 1854 | INV12666 | IBIHM1067-22 | BOLD:AED6294 | OP347210 |
| | <i>Chrysis hydropica</i> Abeille de Perrin, 1878# | INV12735 | IBIHM1136-22 | BOLD:AET2381 | OP347243 |

| Genus | Species | IBI code | BOLD code | BOLD BIN | GenBank |
|----------|--|----------|--------------|------------------------------|--------------------------|
| | <i>Chrysis insperata</i> Chevrier, 1870# | INV12774 | IBIHM1175-22 | BOLD:AET2383 | OP347258 |
| | <i>Chrysis integra</i> Fabricius, 1787# | INV12741 | IBIHM1142-22 | BOLD:AET2382 | OP347285 |
| | <i>Chrysis irreporta</i> Linsenmaier, 1959# | INV12671 | IBIHM1072-22 | BOLD:AER8828 | OP347244 |
| | <i>Chrysis lusitanica</i> Bischoff, 1910 | INV12747 | IBIHM1148-22 | BOLD:ACQ6955 | OP347197 |
| | <i>Chrysis merceti</i> (Trautmann, 1926)# | INV12744 | IBIHM1145-22 | BOLD:AET3720 | OP347260 |
| | <i>Chrysis mixta</i> Dahlbom, 1854# | INV12664 | IBIHM1065-22 | BOLD:AET3717 | OP347207 |
| | <i>Chrysis monticola</i> Linsenmaier, 1999# | INV12681 | IBIHM1082-22 | BOLD:AET3719 | OP347289 |
| | | INV12730 | IBIHM1131-22 | | OP347304 |
| | <i>Chrysis mysticalis</i> Linsenmaier, 1959# | INV12678 | IBIHM1079-22 | BOLD:AET3718 | OP347245 |
| | | INV12736 | IBIHM1137-22 | | OP347271 |
| | <i>Chrysis peninsularis</i> du Buysson, 1887# | INV12667 | IBIHM1068-22 | BOLD:AES0122 | OP347226 |
| | <i>Chrysis crossi</i> Rosa sp. nov.# | INV12727 | IBIHM1128-22 | BOLD:AES0121 | OP347295 |
| | <i>Chrysis pulchella</i> Spinola, 1808 | INV12665 | IBIHM1066-22 | BOLD:AED0619 | OP347198 |
| | <i>Chrysis pulcherrima</i> Lepeletier, 1806# | INV12688 | IBIHM1089-22 | BOLD:AET0271 | OP347172 |
| | <i>Chrysis pyrophana</i> Dahlbom, 1854# | INV12775 | IBIHM1176-22 | BOLD:AET0272 | OP347213 |
| | <i>Chrysis ramburi</i> Dahlbom, 1854 | INV12674 | IBIHM1075-22 | BOLD:AED5814 | OP347263 |
| | <i>Chrysis sculpturata</i> Mocsáry, 1912 | INV12684 | IBIHM1085-22 | BOLD:ABU6373 | OP347310 |
| | <i>Chrysis scutellaris marteni</i> Linsenmaier, 1951 | INV12738 | IBIHM1139-22 | BOLD:ACM0910 | OP347269 |
| | | INV12739 | IBIHM1140-22 | | OP347185 |
| | <i>Chrysis sexdentata</i> Christ, 1791 | INV12783 | IBIHM1184-22 | BOLD:ABU6376 | OP347217 |
| | <i>Chrysis splendidula</i> Rossi, 1790# | INV12740 | IBIHM1141-22 | BOLD:AES6413 | OP347180 |
| | <i>Chrysis subsinuata</i> Marquet, 1879# | INV12662 | IBIHM1063-22 | BOLD:AES4620 | OP347214 |
| | <i>Chrysis varidens</i> Abeille de Perrin, 1878 | INV12668 | IBIHM1069-22 | BOLD:AEE0312 | OP347171 |
| | <i>Chrysis zonata</i> Dahlbom, 1854# | INV12742 | IBIHM1143-22 | BOLD:AET8274 | OP347199 |
| Chrysura | <i>Chrysura austriaca</i> (Fabricius, 1804) | INV12690 | IBIHM1091-22 | BOLD:AAJ3472 | OP347193 |
| | <i>Chrysura cuprea</i> (Rossi, 1790) | INV12692 | IBIHM1093-22 | BOLD:AAP1055 | OP347176 |
| | <i>Chrysura dichroa</i> (Dahlbom, 1854)# | INV12696 | IBIHM1097-22 | BOLD:AET1511 | OP347273 |
| | | INV12776 | IBIHM1177-22 | | OP347215 |
| | <i>Chrysura hybrida</i> (Lepeletier, 1806) | INV12726 | IBIHM1127-22 | BOLD:AAY6924 | OP347177 |

| Genus | Species | IBI code | BOLD code | BOLD BIN | GenBank |
|-------|---|----------|--------------|------------------------------|--------------------------|
| | <i>Chrysura purpureifrons</i> (Abeille de Perrin, 1878) # | INV12694 | IBIHM1095-22 | BOLD:AEU2029 | OP347231 |
| | | INV12693 | IBIHM1094-22 | BOLD:AED1166 | OP347286 |
| | | INV12695 | IBIHM1096-22 | | OP347257 |
| | | INV12849 | IBIHM1250-22 | | OP347296 |
| | | INV12850 | IBIHM1251-22 | | OP347297 |
| | | INV12851 | IBIHM1252-22 | | OP347221 |
| | <i>Chrysura radians</i> (Harris, 1776) | INV12697 | IBIHM1098-22 | BOLD:ABA8702 | OP347225 |
| | <i>Chrysura refulgens</i> (Spinola, 1806) | INV12777 | IBIHM1178-22 | BOLD:ABA7395 | OP347303 |
| | <i>Chrysura rufiventris</i> (Dahlbom, 1854) | INV12699 | IBIHM1100-22 | BOLD:AEC6882 | OP347290 |
| | <i>Chrysura simplex</i> (Dahlbom, 1854) | INV12691 | IBIHM1092-22 | BOLD:AAY6923 | OP347189 |
| | <i>Chrysura sulcata</i> (Dahlbom, 1845) | INV12700 | IBIHM1101-22 | BOLD:ABA7396 | OP347249 |
| | | INV12698 | IBIHM1099-22 | BOLD:AET0222 | OP347261 |
| | | INV12701 | IBIHM1102-22 | | OP347192 |
| | <i>Hedychridium aereolum</i> du Buysson, 1892 | INV12809 | IBIHM1210-22 | BOLD:AAY6930 | OP347195 |
| | <i>Hedychridium anale</i> (Dahlbom, 1854) | INV12717 | IBIHM1118-22 | BOLD:AED4749 | OP347270 |
| | <i>Hedychridium ardens</i> (Coquebert, 1801) | INV12786 | IBIHM1187-22 | BOLD:AAK4640 | OP347236 |
| | <i>Hedychridium buyssoni</i> Abeille de Perrin, 1887# | INV12790 | IBIHM1191-22 | BOLD:AES9011 | OP347196 |
| | <i>Hedychridium caputaureum</i> Trautmann & Trautmann, 1919 | INV12760 | IBIHM1161-22 | BOLD:AAU0775 | OP347183 |
| | <i>Hedychridium valesiense</i> Linsenmaier, 1959# | INV12804 | IBIHM1205-22 | BOLD:AET6828 | OP347275 |
| | <i>Hedychridium chloropygum</i> du Buysson, 1888 | INV12801 | IBIHM1202-22 | BOLD:AAE3258 | OP347187 |
| | | INV12802 | IBIHM1203-22 | | OP347240 |
| | <i>Hedychridium cupratum</i> (Dahlbom, 1854) | INV12806 | IBIHM1207-22 | BOLD:AAY6946 | OP347255 |
| | | INV12807 | IBIHM1208-22 | | OP347252 |
| | <i>Hedychridium cupritibiale</i> Linsenmaier, 1987# | INV12769 | IBIHM1170-22 | BOLD:AES9012 | OP347239 |
| | <i>Hedychridium incrassatum</i> (Dahlbom, 1854) | INV12718 | IBIHM1119-22 | BOLD:AEE0029 | OP347267 |

| Genus | Species | IBI code | BOLD code | BOLD BIN | GenBank |
|---------------------|---|----------|--------------|------------------------------|--------------------------|
| <i>Hedychridium</i> | <i>Hedychridium infans</i> Abeille de Perrin, 1878# | INV12768 | IBIHM1169-22 | BOLD:AES6837 | OP347254 |
| | | INV12781 | IBIHM1182-22 | | OP347191 |
| | <i>Hedychridium jucundum</i> Mocsáry, 1889# | INV12784 | IBIHM1185-22 | BOLD:AES6836 | OP347262 |
| | | INV12785 | IBIHM1186-22 | | OP347232 |
| | | INV12810 | IBIHM1211-22 | BOLD:AES6835 | OP347170 |
| | | INV12811 | IBIHM1212-22 | | OP347256 |
| | | INV12764 | IBIHM1165-22 | BOLD:AAZ0056 | OP347237 |
| | <i>Hedychridium mediocrum</i> Linsenmaier, 1987 | INV12716 | IBIHM1117-22 | BOLD:AAE3260 | OP347279 |
| | | INV12803 | IBIHM1204-22 | | OP347308 |
| | <i>Hedychridium monochroum</i> du Buysson, 1888 | INV12800 | IBIHM1201-22 | BOLD:AAY1978 | OP347206 |
| | <i>Hedychridium reticulatum</i> Abeille de Perrin, 1878# | INV12763 | IBIHM1164-22 | BOLD:AER9655 | OP347248 |
| | | INV12791 | IBIHM1192-22 | | OP347203 |
| | | INV12792 | IBIHM1193-22 | | OP347282 |
| | | INV12793 | IBIHM1194-22 | | OP347301 |
| | <i>Hedychridium roseum</i> (Rossi, 1790) | INV12805 | IBIHM1206-22 | BOLD:AAE3259 | OP347281 |
| | <i>Hedychridium sculpturatum</i> Abeille de Perrin, 1877# | INV12812 | IBIHM1213-22 | BOLD:AET6828 | OP347181 |
| | | INV12813 | IBIHM1214-22 | BOLD:AAE3258 | OP347276 |
| | <i>Hedychridium scutellare</i> (Tournier, 1878)# | INV12761 | IBIHM1162-22 | BOLD:AES0428 | OP347291 |
| | | INV12788 | IBIHM1189-22 | | OP347175 |
| | <i>Hedychridium sevillanum</i> Linsenmaier, 1968# | INV12765 | IBIHM1166-22 | BOLD:AET6827 | OP347242 |
| | | INV12767 | IBIHM1168-22 | BOLD:AES0429 | OP347259 |
| | | INV12789 | IBIHM1190-22 | | OP347223 |
| | <i>Hedychridium calcarium</i> Rosa sp. nov.# | INV12794 | IBIHM1195-22 | BOLD:AET9698 | OP347204 |
| | | INV12795 | IBIHM1196-22 | | OP347307 |
| | | INV12796 | IBIHM1197-22 | | OP347169 |
| | <i>Hedychridium subroseum prochloropygum</i> Linsenmaier, 1959# | INV12715 | IBIHM1116-22 | BOLD:AES2437 | OP347209 |
| | <i>Hedychridium vachali</i> Mercet, 1915# | INV12766 | IBIHM1167-22 | BOLD:AET6826 | OP347280 |
| <i>Hedychrum</i> | <i>Hedychrum longicolle</i> Abeille de Perrin, 1877 | INV12723 | IBIHM1124-22 | BOLD:AED0972 | OP347298 |

| Genus | Species | IBI code | BOLD code | BOLD BIN | GenBank |
|--------------------|--|----------|--------------|------------------------------|--------------------------|
| | <i>Hedychrum micans europaeum</i> Linsenmaier, 1959 | INV12724 | IBIHM1125-22 | BOLD:AAK4644 | OP347194 |
| | <i>Hedychrum niemelai</i> Linsenmaier, 1959 | INV12721 | IBIHM1122-22 | BOLD:AAU1294 | OP347300 |
| | <i>Hedychrum nobile</i> (Scopoli, 1763) | INV12722 | IBIHM1123-22 | BOLD:AAK4644 | OP347266 |
| | | INV12771 | IBIHM1172-22 | | OP347168 |
| | <i>Hedychrum rutilans</i> Dahlbom, 1854 | INV12720 | IBIHM1121-22 | BOLD:AAM3491 | OP347220 |
| | <i>Hedychrum viridiaureum</i> Tournier, 1877 | INV12719 | IBIHM1120-22 | BOLD:AAM3491 | OP347294 |
| | | INV12770 | IBIHM1171-22 | BOLD:AAK4643 | OP347247 |
| | | INV12782 | IBIHM1183-22 | BOLD:AAM3491 | OP347174 |
| <i>Holopyga</i> | <i>Holopyga calida</i> Linsenmaier, 1951# | INV12705 | IBIHM1106-22 | | OP347202 |
| | | INV12754 | IBIHM1155-22 | BOLD:AES0090 | OP347227 |
| | <i>Holopyga lucida</i> (Lepeletier, 1806) # | INV12708 | IBIHM1109-22 | BOLD:AES0091 | OP347182 |
| | <i>Holopyga fastuosa</i> (Lucas, 1849) | INV12709 | IBIHM1110-22 | BOLD:AAZ6194 | OP347292 |
| | | INV12710 | IBIHM1111-22 | | OP347190 |
| | | INV12780 | IBIHM1181-22 | | OP347253 |
| | | INV12753 | IBIHM1154-22 | BOLD:AAY6928 | OP347216 |
| | <i>Holopyga fervida</i> (Fabricius, 1781) | INV12706 | IBIHM1107-22 | BOLD:ACV6331 | OP347208 |
| | | INV12757 | IBIHM1158-22 | BOLD:AAY9735 | OP347287 |
| | <i>Holopyga generosa</i> (Förster, 1853) | INV12712 | IBIHM1113-22 | BOLD:AAZ6194 | OP347288 |
| | | INV12713 | IBIHM1114-22 | | OP347201 |
| | <i>Holopyga inflammata</i> (Förster, 1853) # | INV12759 | IBIHM1160-22 | BOLD:AET1451 | OP347284 |
| | <i>Holopyga jurinei</i> Chevrier, 1862# | INV12714 | IBIHM1115-22 | | OP347233 |
| | | INV12756 | IBIHM1157-22 | BOLD:AET1450 | OP347235 |
| | <i>Holopyga similis</i> Mocsáry, 1889 | INV12787 | IBIHM1188-22 | BOLD:AED0274 | OP347178 |
| | <i>Holopyga merceti</i> Kimsey, 1991# | INV12755 | IBIHM1156-22 | BOLD:AES1216 | OP347186 |
| <i>Parnopes</i> | <i>Parnopes</i> sp.# | INV12779 | IBIHM1180-22 | BOLD:AET2814 | OP347218 |
| <i>Philoctetes</i> | <i>Philoctetes abeillei</i> du Buysson, 1892# | INV12704 | IBIHM1105-22 | BOLD:AEU5026 | OP347179 |
| | <i>Philoctetes punctulatus</i> (Dahlbom, 1854) # | INV12703 | IBIHM1104-22 | BOLD:AEU5027 | OP347264 |
| | | INV12749 | IBIHM1150-22 | | OP347272 |
| | | INV12750 | IBIHM1151-22 | | OP347238 |

| Genus | Species | IBI code | BOLD code | BOLD BIN | GenBank |
|----------------------|---|----------|--------------|------------------------------|--------------------------|
| <i>Pseudochrysis</i> | <i>Pseudochrysis humboldti</i> (Dahlbom, 1845) # | INV12659 | IBIHM1060-22 | BOLD:AEU3425 | OP347246 |
| | <i>Pseudochrysis incrassata</i> (Spinola, 1838) # | INV12660 | IBIHM1061-22 | BOLD:AEU3426 | OP347278 |
| | <i>Pseudomalus auratus</i> (Linnaeus, 1758) | INV12751 | IBIHM1152-22 | BOLD:AAH8217 | OP347251 |
| | <i>Pseudomalus violaceus</i> (Scopoli, 1763) | INV12752 | IBIHM1153-22 | BOLD:ABX9998 | OP347299 |
| <i>Spintharina</i> | <i>Spintharina cuprata</i> (Dahlbom, 1854) # | INV12657 | IBIHM1058-22 | BOLD:AET6497 | OP347277 |
| | <i>Spintharina versicolor</i> (Spinola, 1808) | INV12656 | IBIHM1057-22 | BOLD:AAJ3630 | OP347224 |
| <i>Stilbum</i> | <i>Stilbum westermanni</i> Dahlbom, 1845 # | INV12654 | IBIHM1055-22 | BOLD:AES3895 | OP347311 |
| | | INV12655 | IBIHM1056-22 | | OP347306 |
| | <i>Stilbum cyanurum</i> (Forster, 1771) | INV12725 | IBIHM1126-22 | BOLD:AAJ4180 | OP347184 |
| <i>Trichrysis</i> | <i>Trichrysis cyanea</i> (Linnaeus, 1758) | INV12658 | IBIHM1059-22 | BOLD:AAH7935 | OP347188 |

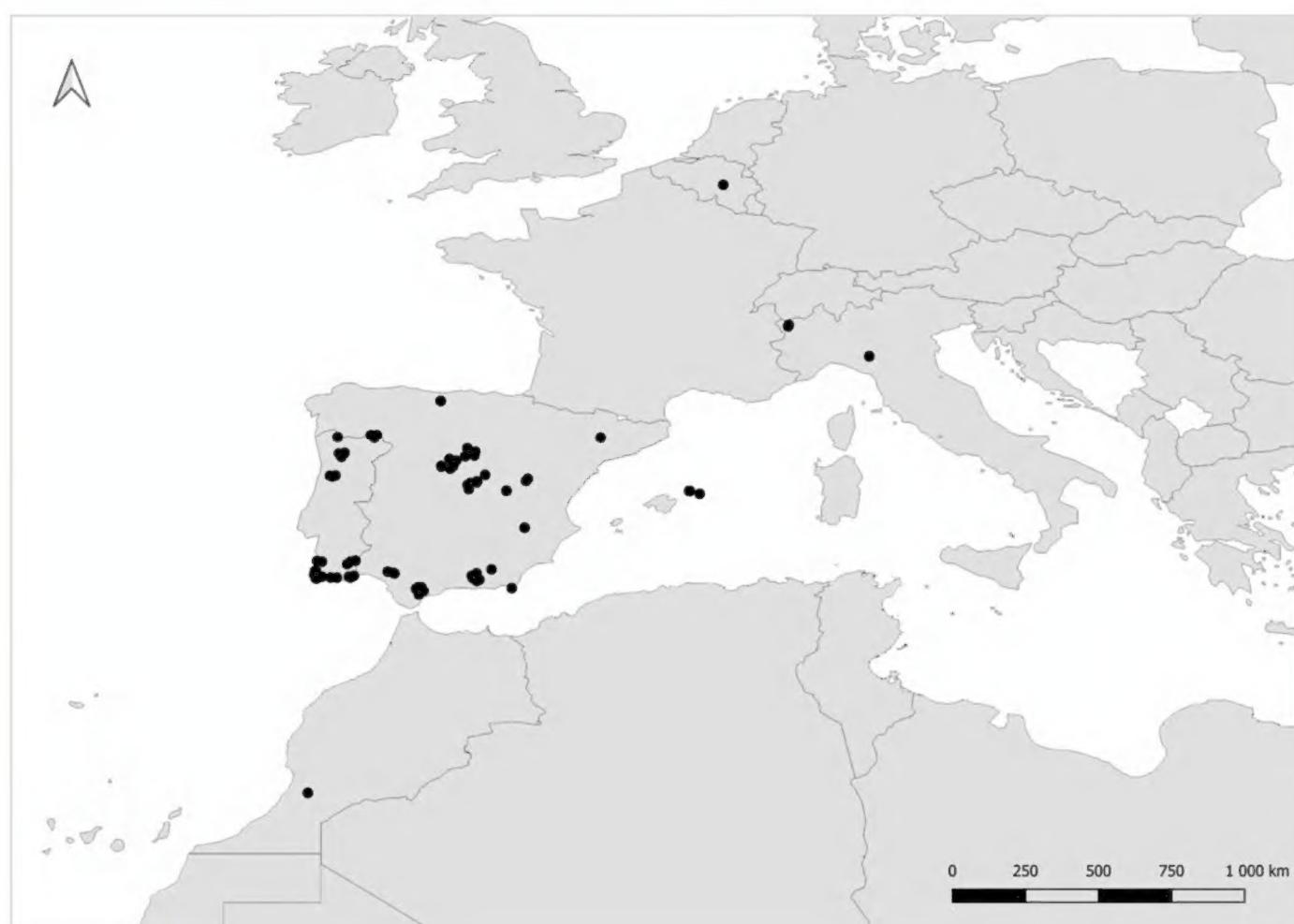


Figure 1. [doi](#)

Map of the localities where cuckoo wasps samples were collected.

Specimens were captured with an entomological net, euthanised by exposure to ethyl acetate and pinned and dried within 24 hours to achieve maximum suitability for DNA extraction and amplification.

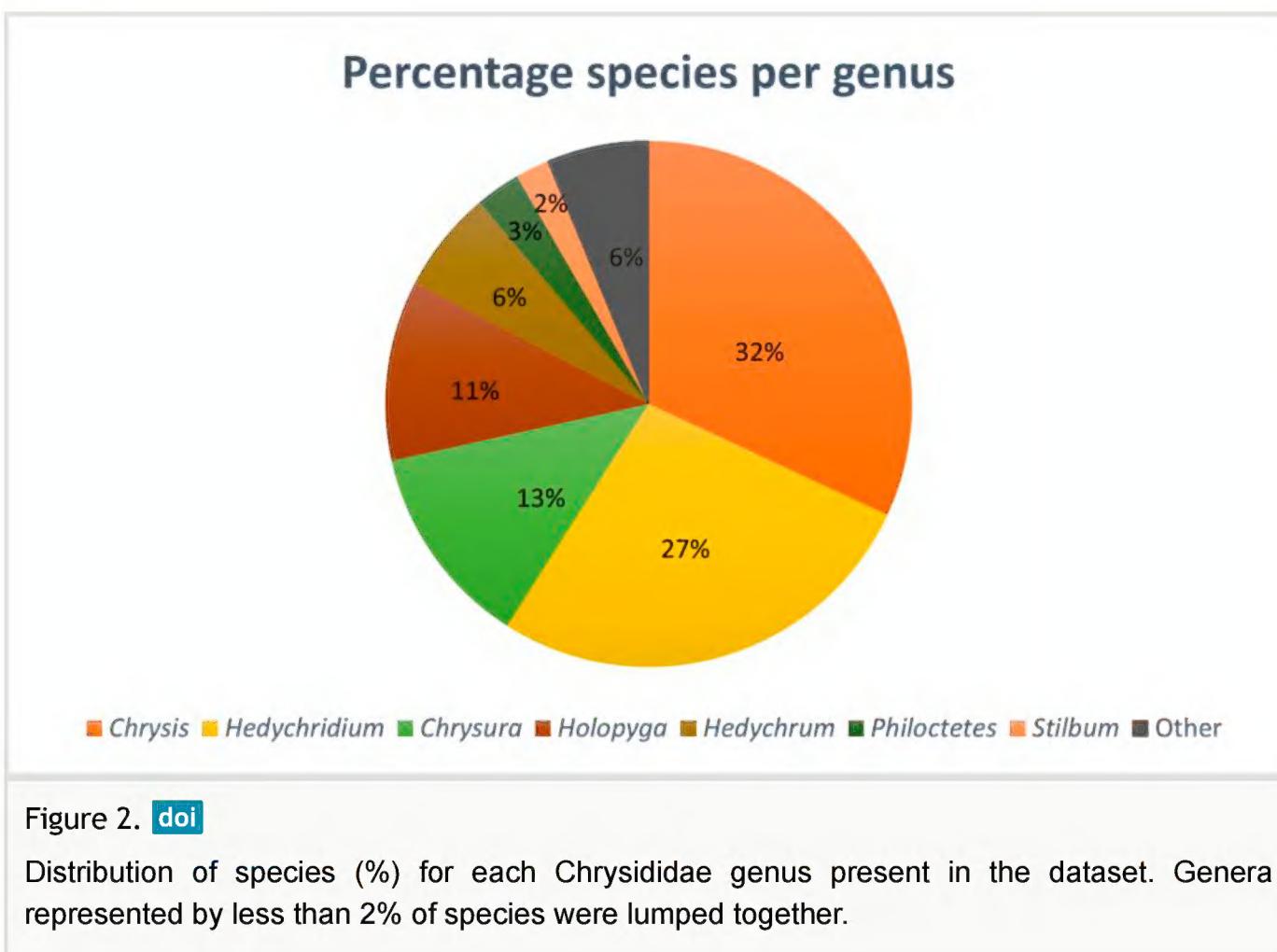


Figure 2. [doi](#)

Distribution of species (%) for each Chrysididae genus present in the dataset. Genera represented by less than 2% of species were lumped together.

DNA was extracted using a QIAamp DNA Micro Kit that is designed to extract higher concentrations of genetic material from samples with small amounts of DNA. DNA amplification was performed using two different primer pairs, that amplify partially overlapping fragments (LC + BH) of the 658 bp barcoding region of the COI mitochondrial gene (Folmer et al. 1994). We used the primers FwhF1 (Vamos et al. 2017) + C_R (Shokralla et al. 2015) for LC and BF3 (Elbrecht et al. 2019) + BR2 (Elbrecht and Leese 2017) for BH amplification, all modified with Illumina adaptors. PCRs were performed in 10 μ l reactions, containing 5 μ l of Multiplex PCR Master Mix (Qiagen, Germany), 0.3 μ l of each 10 mM primer and 1-2 μ l of DNA, with the remaining volume in water. PCR cycling conditions consisted in an initial denaturation at 95°C for 15 min, followed by 45 cycles of denaturation at 95°C for 30 sec, annealing at 45°C for 45 sec and extension at 72°C for 45 sec, with a final elongation step at 60°C for 10 min. All DNA extracts were deposited in the IBI collection.

Successful amplification was validated through 2% agarose gel electrophoresis and samples selected for sequencing followed for a second PCR, where Illumina P5 and P7 adapters with custom 7 bp long barcodes were attached to each PCR product. The index PCR was performed in a volume of 10 μ l, including 5 μ l of KAPA HiFi PCR Kit (KAPA Biosystems, U.S.A.), 0.5 μ l of each 10 mM indexing primer and 2 μ l of diluted PCR product (usually 1:4). PCR cycling conditions were as before, except that only 10 cycles were performed and at an annealing temperature of 55°C. The amplicons were purified using AMPure XP beads (New England Biolabs, U.S.A.) and quantified using NanoDrop 1000 (Thermo Scientific, U.S.A.). Clean PCR products were then pooled equimolarly per fragment. Each pool was quantified with KAPA Library Quantification Kit Illumina®

Platforms (KAPA Biosystems, U.S.A.) and the 2200 Tapestation System (Agilent Technologies, California, USA) was used for fragment length analysis prior to sequencing (Paupério et al. 2018). DNA sequencing was done at CIBIO facilities on an Illumina MiSeq benchtop system, using a V2 MiSeq sequencing kit (2x 250 bp).

Illumina sequencing reads were processed using OBITools (Boyer et al. 2016) and VSEARCH (Rognes et al. 2016). Briefly, paired-end reads were aligned, collapsed into exact sequence variants, filtered by length, denoised and checked for chimeras. The resulting sequences from both LC and BH fragments of each sample were further assembled using CAP3 (Huang and Madan 1999) to produce a single 658 bp contig per sample.

All sequences in the dataset were submitted to BOLD and GenBank databases and, to each sequenced specimen, the morphological identification was contrasted with the results of the BLAST of the newly-generated DNA barcodes in the BOLDIdentification Engine. In order to clarify the taxonomic status of problematic groups, DNA barcodes generated here were analysed with other sequences from across the West Palaearctic downloaded from BOLD and GenBank. Sequences (658 bp) were aligned using SeaView (Gouy et al. 2010) and a neighbour-joining phylogeny was run with 10,000 bootstraps. Intra- and interspecific distances were calculated using MEGA-X (Kumar et al. 2018).

Acronyms and abbreviations

NMLU = Natur-Museum, Lucerne, Switzerland

PRC = Paolo Rosa Collection, Bernareggio, Italy

TWC = Thomas J. Wood Collection, Mons, Belgium

ICC = Ian Cross Collection, Briantspuddle, Dorset, United Kingdom

In the text, the following abbreviations are used for morphological terms:

- **F1, F2, F3** = flagellomeres 1, 2, 3
- **MOD** = anterior ocellar diameter
- **MS** = malar space, the shortest distance between base of mandible and margin of compound eye
- **OOL** = oculo-ocellar line, the shortest distance between lateral ocellus and compound eye
- **P** = pedicel
- **PD** = puncture diameter
- **POL** = the shortest distance between posterior ocelli
- **T1–T3** = metasomal terga 1 to 3
- **vs.** = versus

Photographs were taken with a Camera Olympus E-M1 Mark II with the Olympus Zuiko 60 mm objective and stacked with the software Helicon Focus (ver. 7.6). Further image processing was completed with Adobe Photoshop CS6.0.

Data resources

The InBIO Barcoding Initiative Database: contribution to the knowledge on DNA barcodes of "European Chrysidae" dataset can be downloaded from the Public Data Portal of BOLD (<http://dx.doi.org/10.5883/DS-IBIHY02>) in different formats (data as dwc, xml or tsv and sequences as fasta files). Alternatively, BOLD users can log-in and access the dataset via the Workbench platform of BOLD. All records are also searchable within BOLD, using the search function of the database.

The version of the dataset, at the time of writing the manuscript, is included as Suppl. material 1 in the form of one text file with specimen data information, as Suppl. material 2 in the form of DWC file specimen data and one fasta file containing all sequences as downloaded from BOLD (Suppl. material 3).

Taxon treatments

Hedychridium calcarium Rosa, sp. n.

- ZooBank [3F4762B4-7EAB-4B8F-9B63-F18EA15255C4](http://zoobank.org/3F4762B4-7EAB-4B8F-9B63-F18EA15255C4)

Materials

Holotype:

- scientificName: *Hedychridium* sp.; phylum: Arthropoda; class: Insecta; order: Hymenoptera; family: Chrysidae; genus: *Hedychridium*; country: Spain; locality: Sierra de Baza, Prados del Rey; decimalLatitude: 37.375; decimalLongitude: -2.854; year: 2021; month: 6; day: 25; individualID: INV12794; individualCount: 1; lifeStage: Adult; catalogNumber: INV12794; recordNumber: INV12794; recordedBy: Thomas Wood; otherCatalogNumbers: IBIHM1195-22; identifiedBy: Paolo Rosa; institutionCode: Universite de Mons; occurrenceID: 0D71E997-8420-5092-B584-2E6619F2EA61

Paratypes:

- scientificName: *Hedychridium* sp.; phylum: Arthropoda; class: Insecta; order: Hymenoptera; family: Chrysidae; genus: *Hedychridium*; country: Spain; locality: Noguera de Albarracin, Barranco de la Olmeda; decimalLatitude: 40.462; decimalLongitude: -1.614; year: 2021; month: 6; day: 27; individualID: INV12795; individualCount: 1; lifeStage: Adult; catalogNumber: INV12795; recordNumber: INV12795; recordedBy: Thomas Wood; otherCatalogNumbers: IBIHM1196-22; identifiedBy: Paolo Rosa; institutionCode: Universite de Mons; occurrenceID: DF256B6E-111E-51B6-8356-0E121E5033F4
- scientificName: *Hedychridium* sp.; phylum: Arthropoda; class: Insecta; order: Hymenoptera; family: Chrysidae; genus: *Hedychridium*; country: Spain; locality: Villar del Cobo, Barranco de los Oncenachos; decimalLatitude: 40.397; decimalLongitude:

-1.674; year: 2021; month: 6; day: 19; individualID: INV12796; individualCount: 1; lifeStage: Adult; catalogNumber: INV12796; recordNumber: INV12796; recordedBy: Thomas Wood; otherCatalogNumbers: IBIHM1197-22; identifiedBy: Paolo Rosa; institutionCode: Universite de Mons; occurrenceID: D746B3CE-8112-585E-B5AA-FBB5866002CB

Description

Female. Body length 5.0–5.4 mm (holotype 5.4 mm (Fig. 3)). Forewing length 3.0–3.5 mm.

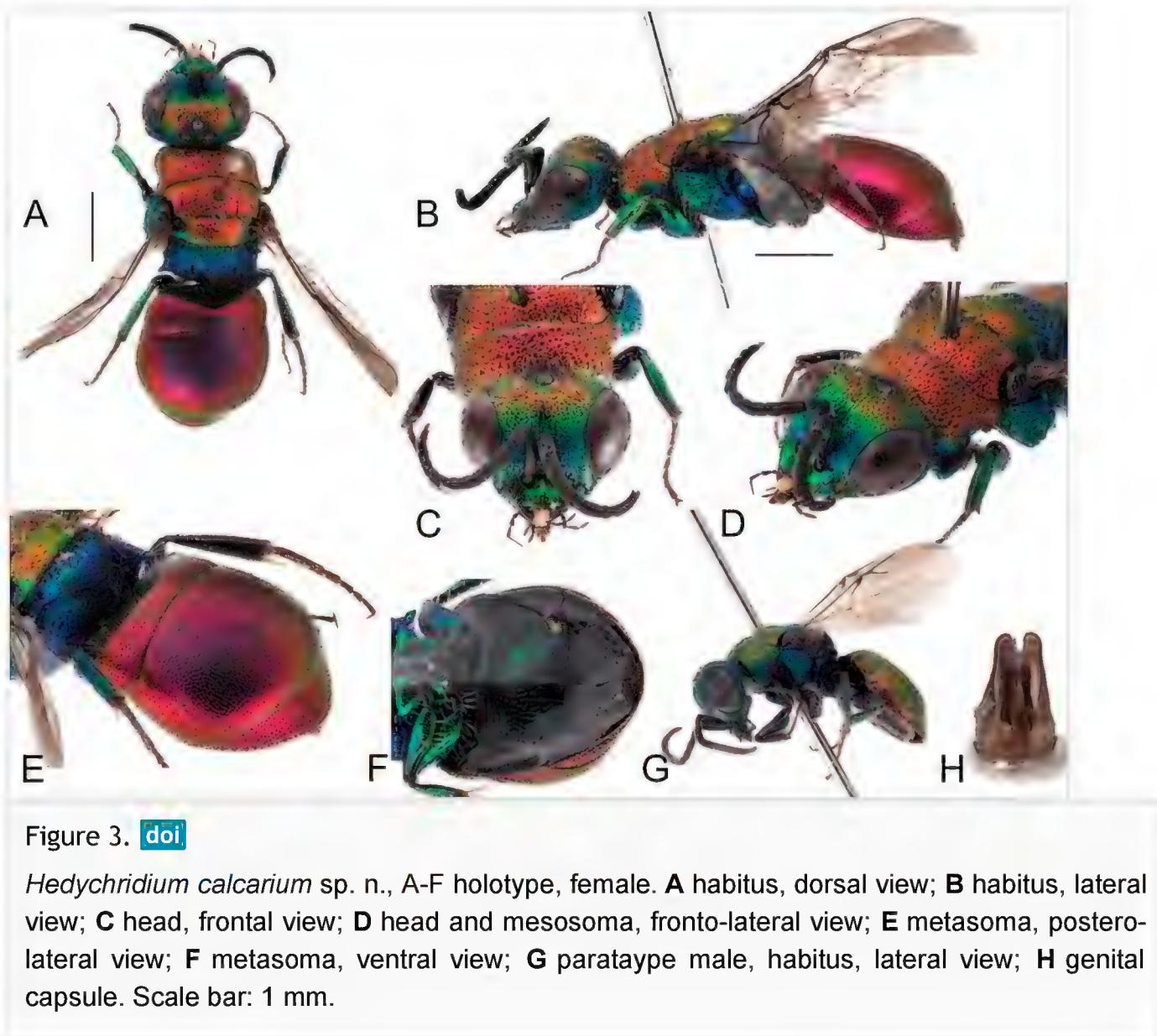


Figure 3. [doi](#)

Hedychridium calcarium sp. n., A-F holotype, female. **A** habitus, dorsal view; **B** habitus, lateral view; **C** head, frontal view; **D** head and mesosoma, fronto-lateral view; **E** metasoma, postero-lateral view; **F** metasoma, ventral view; **G** paratype male, habitus, lateral view; **H** genital capsule. Scale bar: 1 mm.

Head. Brow with medium, contiguous punctures (ca. $0.4 \times$ MOD), suddenly decreasing diameter from frontal declivity to malar spaces and clypeus (Fig. 3C); face, in frontal view, micropunctate along inner eye margin; scapal basin with polished intervals; medial line complete from anterior ocellus to clypeus; clypeus finely punctate with wide polished intervals; clypeal apical margin thickened, triangularly-shaped, non-metallic brown; ocellar area with small punctures, without line connecting posterior ocelli; temples regularly rounded, double punctate. OOL = $1.9 \times$ MOD; POL = $1.5 \times$ MOD; MS = $0.6 \times$ MOD; relative length of P:F1:F2:F3 = 1:1.1:0.8:0.8. Malar space as long as antennal thickness.

Mesosoma. Pronotum with punctuation irregularly sized, mostly contiguous and large, umbelicate punctures up to 0.6 MOD; intervals between large punctures densely micropunctate. Mesoscutum with similar punctuation, yet punctures relatively smaller and micropunctures sparser compared to intervals on pronotum. Scutellum with polished intervals and sparse micropunctures. Metascutellum with reticulate-foveate punctures ($0.8 \times$ MOD). Metapectal-propodeal complex with metapostnotum wider than in other species (Fig. 4A) and posterior propodeal projection [= propodeal teeth] triangular, with thickened, blunt apex, slightly pointing backwards. Forewing medial vein 1.5 times as long as RS stub, medially gently arched; Rs stub as long as pterostigma. Hind leg unmodified, metatibia entirely black, without visible spots or depressions.

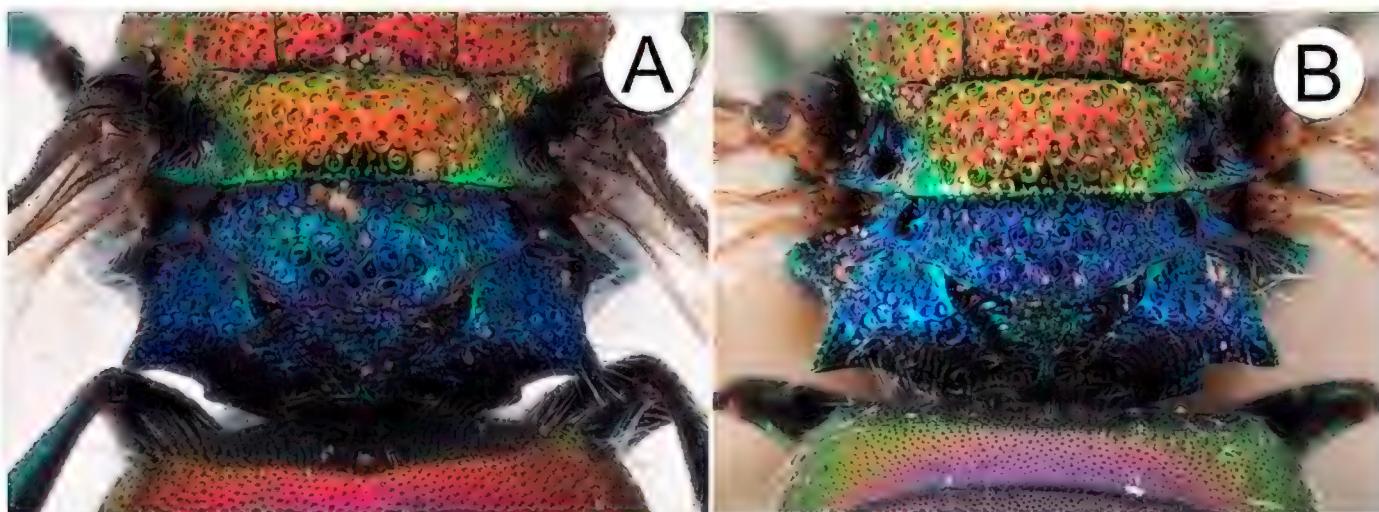


Figure 4. [doi](#)

Scutellum, metanotum, metapectal-propodeal complex **A** *Hedyphridium calcarium* sp. nov., holotype, female; **B** *Hedyphridium jucundum*, female, from Italy (PRC).

Metasoma. Punctuation on terga minute, even, sparse, regularly spaced, 1–2 PD apart. Third tergum laterally with denser, deeper punctures (Fig. 3E); posterior margin with hyaline rim (2 PD).

Colouration. Head blue with two large golden-red spots on brow, between anterior ocellus and eyes; clypeus, malar space and base of mandible greenish; ocelli area blackish. Pronotum and mesonotum red, lateral and posterior margin of scutellum green; rest of mesosoma blue, with green mesopleuron and legs. Metasoma dorsally red, ventrally black with two large, oblique green to blue spots on second sternum (Fig. 3C). Mandible entirely dark brown. Scape black with slight metallic reflection, pedicel and flagellomeres black; tegula black. Wings slightly infuscate.

Male: Paratype from Teruel similar to female, with face laterally covered with appressed, silvery setae; antennae elongate, with slender flagellum and cylindrical articles. Paratype from Granada smaller (4.0 mm) with red colouration turned to green (Fig. 3G); genital capsule as in Fig. 3H, with slender cuspis, apically unmodified.

Diagnosis

The genus *Hedychridium* Abeille de Perrin, 1878 in Iberia includes 34 species and two subspecies (Rosa and Soon 2012), whereas three previous members were recently moved from the genus *Hedychridium* to the genus *Colopopyga* Semenov-Tian-Shanskji, 1954 (Rosa 2017). Twenty-five of these species are known from Portugal (Rosa et al., in preparation). Mingo (1994) listed only 26 species for Iberia, another nine were overlooked, but has previously been described or cited from Spain by Linsenmaier (Linsenmaier 1959, Linsenmaier 1968, Linsenmaier 1987) and two species, *H. infantum* Linsenmaier, 1997 and *H. balearicum* Strumia, 2013, were described later.

The list of *Hedychridium* species known from Iberia is given below, with species subdivided by species groups following Linsenmaier's classification (Rosa et al. 2022) and modifications proposed by Pauli et al. (2019), based on multigene molecular analyses.

anale group: *Hedychridium anale* (Dahlbom, 1854); *H. dubium* Mercet, 1904;

ardens group: *H. adventicum* Zimmermann, 1961, *H. aereolum* du Buysson, 1891, *H. ardens* (Coquebert, 1801), *H. buyssoni* Abeille de Perrin, 1887, *H. cupritibiale* Linsenmaier, 1987, *H. ibericum* Linsenmaier, 1959, *H. infans* Abeille de Perrin, 1879, *H. infans santschii* Trautmann, 1927, *H. infantum* Linsenmaier, 1987, *H. jucundum* (Mocsáry, 1889), *H. marteni* Linsenmaier, 1951; *H. reticulatum* Abeille de Perrin, 1878, *H. sevillanum* Linsenmaier, 1968;

coriaceum group: *H. coriaceum* (Dahlbom, 1854), *H. krajniki* Balthasar, 1953;

cupratum group: *H. cupratum* (Dahlbom, 1854);

femoratum group: *H. elegantulum* du Buysson, 1887, *H. femoratum* (Dahlbom, 1854), *H. gratiosum* Abeille de Perrin, 1878;

heliophilum group: *H. heliophilum* du Buysson, 1887, *H. vachali* Mercet, 1915;

incrassatum group: *H. incrassatum* (Dahlbom, 1854);

monochroum group: *H. balearicum* Strumia, 2013, *H. carmelitanum* Mercet, 1915, *H. minutissimum* Mercet, 1915, *H. monochroum* du Buysson, 1888;

plagiatum group: *H. andalusicum* Trautmann, 1920, *H. franciscanum* Linsenmaier, 1987;

roseum group: *H. chloropygum* du Buysson, 1888, *H. mediocrum* Linsenmaier, 1987, *H. roseum* (Rossi, 1790), *H. scutellare* (Tournier, 1878), *H. subroseum* Linsenmaier, 1959, *H. subroseum prochloropygum* Linsenmaier, 1959.

The following three species were moved in the genus *Colopopyga*, supported by morphological and molecular evidence (Rosa 2017, Pauli et al. 2019): *C. auriventris*

(Mercet, 1904), *C. flavipes* (Eversmann, 1858) and *C. temperata* (Lisenmaier, 1959). Three species are also known for the Canary islands and are considered endemic: *Hedychridium extraneum* Lisenmaier, 1993, *H. tricavatum* Lisenmaier, 1993 and *H. viridicupreum* Lisenmaier, 1993.

Another species, *Hedychridium suave* (Tournier, 1878), was described from Spain (Andalucía) and has been considered to be a synonym of *H. roseum* by Lisenmaier (1951), Mingo (1994) and Kimsey and Bohart (1991), who erroneously placed the type locality in Switzerland (Léman area). None of these authors examined the type deposited at the Museum in Geneva. According to the labels pinned with the type specimen, the type locality is Tangier in Morocco and not Andalucía. *Hedychridium suave* does not belong to the *roseum* group, but to the *femoratum* group; it is a valid species and, based on its aspect and colouration, this taxon should be a North African species and the Andalusian locality is an error. Tournier is well-known for confusing European and Moroccan localities, as has already happened in other insect families as well as in Chrysididae (see the case of *Chrysis superba* Tournier, 1879 in Lisenmaier (1968). For the moment, we do not consider *H. suave* to be a member of the Iberian fauna.

Hedychridium calcarium sp. nov. belongs to the *ardens* species group due to the shape of the second metatarsomere which is longer than the third, the punctate scapal basin, the general habitus and the body colouration (Fig. 3). *Hedychridium calcarium* sp. nov. has small to medium dimensions, from 4.0 to 5.4 mm; head blue with two red patches on brow between anterior ocellus and compound eye; black ocellar area; red pronotum and mesonotum, rest of mesosoma blue with greenish reflections; metasoma dorsally red and ventrally black with two large and oblique green-bluish spots on the second sternum. Punctuation dense, even and deep on vertex; the largest punctures deep and umbilicate on pronotum, with intervals densely micropunctate; mesoscutum with smaller, shallower and sparser punctures, intervals less densely micropunctate compared to pronotum; metanotum with sparse micropunctures on shining intervals; metapostnotum distinctly enlarged compared to the same morphological part of the closest species, *H. jucundum* (Fig. 4), in which it is triangular. Metasomal sculpture with even, dense and small punctures equally spaced; apical margin of the third tergum with wide hyaline margin (2–3 PD).

Besides different body sculpture and morphological characters, *Hedychridium calcarium* sp. nov. can be immediately separated from *H. ardens*, *H. marteni* and *H. ibericum* by its blue metanotum, contrasting with red scutellum (concolourous in the other species); from *H. cupritibiale* by the blue face, contrasting with the red head on vertex (entirely red in *H. sevillanum*); from *H. sevillanum* by the different body colour, which is green to bronze in the latter and by the metanotum bronze to green, slightly contrasting with the rest of the red body colour. For comparison, pictures of *H. ardens* can be found in Paukkunen et al. (2015) and pictures of all the remaining species can be found in the illustrated catalogue of Lisenmaier's types (Rosa et al. 2022). Finally, *H. infans*, *H. adventicium* and *H. infantum* can be immediately separated by their very small size (2–3 mm) and the different colouration, the first having metallic tegulae (a

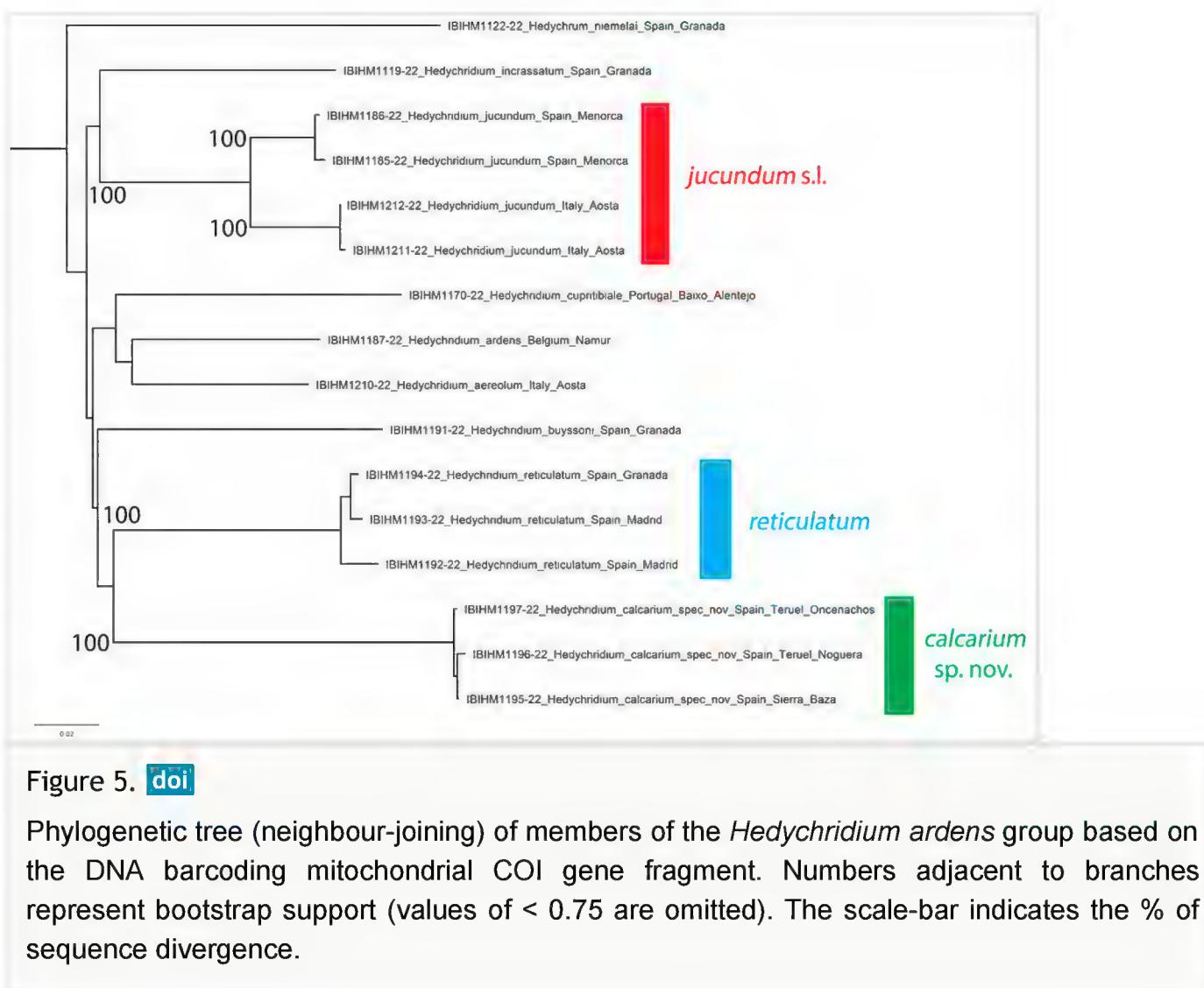
unique feature), the other two a green line along the posterior margin of the pronotum. The species morphologically and chromatically closer to *H. calcarium* sp. nov. are *H. jucundum*, *H. reticulatum* sensu Lisenmaier (1959) and *H. buyssoni*. However, *H. jucundum* can be differentiated by a dark to black spot on discum of second tergum and by the vertex entirely golden to red; in case of doubt, the triangular shape of the metapostnotum is diagnostic (Fig. 4); *H. reticulatum* by the red mesopleuron and, finally, *H. buyssoni* by the green vertex, the stocky body, the first tergum shorter medially and angled on anterior margins, the metasoma with denser and deeper punctures. The male of *H. calcarium* sp. nov. has the same colouration of the female and can be separated from the similar male of *H. jucundum* by the colour of the head and the shape of the genital capsule with cuspis apically slender, unmodified (vs. apically enlarged and curved in *H. jucundum* (see Rosa 2017)).

Outside the Iberian Peninsula, only *Hedychridium bytinskii* Lisenmaier, 1959 can be confused with *H. calcarium* sp.nov. *H. bytinskii* was described from Palestine and is known from Greece and Turkey (Lisenmaier 1968, Lisenmaier 1999). Lisenmaier (Lisenmaier 1968, Lisenmaier 1999) listed this species from Morocco, but the Moroccan specimens may actually belong to the western Mediterranean species *H. calcarium*. The latter can be immediately recognised by the dark metasomal sterna, with two small dark green spots on the second sternum, whereas *H. bytinskii* specimens from the east Mediterranean have the first sternum (largely) and second sternum (entirely) bright green (see pictures of the type in Rosa et al. (2022), Fig. 7E). The second sternum of *H. bytinskii* is also characterised by only a few and sparse punctures bearing long setae, whereas *H. calcarium* has a denser punctuation (Fig. 1F) with short setae that are approximately one third as long as those of *H. bytinskii*. The colour pattern of the head also differs between the two species, with the entire vertex in *H. bytinskii* coloured flame red, distinctly contrasting the blue head and the green declivity of the frons, with the ocelli area flame red; the red area on the vertex of *H. calcarium* is less strongly contrasting and the ocelli area is black. The scutellum is entirely flame red in *H. bytinskii*, whereas it is metallic green on its posterior margin in *H. calcarium*. Barcoding analyses of the eastern Mediterranean *H. bytinskii* are needed to evaluate the genetic distance between the two species.

Genetics: *Hedychridium calcarium* sp. nov. is very distinct genetically (Fig. 5), showing a low average intraspecific genetic distance of 0.30%. It is strongly separated from the nearest relative in the phylogenetic tree *H. reticulatum* by an average genetic distance of 16.45% (range 16.11-16.72%). By direct genetic distance, it is closest to *H. jucundum* specimens from Italy, separated by an average of 16.13% (range 15.96-16.41%). As a note, *H. jucundum* specimens from Italy are separated from *H. jucundum* specimens from Menorca by an average of 4.94% (range 4.71-5.17%). This requires further investigation.

Etymology

The epithet *calcarium* derives from the Latin adjective *calcarius* related to the limestone habitat of the species.



Distribution

Spain (provinces of Teruel and Granada). At each locality, the species was found in dry grassland on calcareous soil, such as at the Barranco de los Oncenachos (Fig. 6).



Figure 6. [doi](#)

Villar del Cobo, Barranco de los Oncenachos, province of Teruel, Spain. Collecting site for *Hedychridium calcarium* Rosa sp. nov.

Ecology

The host is unknown, but is likely to be a small apoid wasp, in line with other members of the *Hedychridium ardens* group.

***Chrysis crossi* Rosa, sp. n.**

- ZooBank [759C3BE4-F1BE-47B2-9C43-0FFD17612E43](https://doi.org/10.1544/759C3BE4-F1BE-47B2-9C43-0FFD17612E43)

Materials

Holotype:

- a. phylum: Arthropoda; class: Insecta; order: Hymenoptera; family: Chrysididae; genus: *Chrysis*; specificEpithet: *crossi*; country: Portugal; locality: Salema; decimalLatitude: 37.06; decimalLongitude: -8.83; year: 2017; month: 4; day: 16; individualID: INV12727; individualCount: 1; sex: F; lifeStage: Adult; catalogNumber: INV12727; recordNumber: INV12727; recordedBy: I. C. Cross; otherCatalogNumbers: IBIHM1128-22; identifiedBy: Paolo Rosa; institutionCode: Universite de Mons; occurrenceID: 6CF2BF38-8154-53D2-AB88-640F8CDD5821

Paratypes:

- a. phylum: Arthropoda; class: Insecta; order: Hymenoptera; family: Chrysididae; genus: *Chrysis*; specificEpithet: *crossi*; country: Portugal; locality: 3 km N Mexilhoeira Grande, Poio; decimalLatitude: 37.2; decimalLongitude: -8.6; year: 2005; month: 4; day: 30; individualCount: 1; sex: M; lifeStage: Adult; recordedBy: J. Smit (JSC); identifiedBy: Paolo Rosa; occurrenceID: 9849F20C-7561-5544-B4E2-430469C8A89F
- b. phylum: Arthropoda; class: Insecta; order: Hymenoptera; family: Chrysididae; genus: *Chrysis*; specificEpithet: *crossi*; country: Portugal; locality: Carrapateira, Praia da Bordeira; decimalLatitude: 37.2; decimalLongitude: -8.9; year: 2006; month: 4; day: 16; individualCount: 2; sex: M; lifeStage: Adult; recordedBy: M. & E. Howe (PRC); identifiedBy: Paolo Rosa; occurrenceID: 01CC81B6-FEC3-5EBD-B89E-B7195661A5AD
- c. phylum: Arthropoda; class: Insecta; order: Hymenoptera; family: Chrysididae; genus: *Chrysis*; specificEpithet: *crossi*; country: Portugal; locality: Carrapateira; decimalLatitude: 37.2; decimalLongitude: -8.9; year: 2016; month: 4; day: 26; individualCount: 1; sex: M; lifeStage: Adult; recordedBy: I.C. Cross (ICC); identifiedBy: Paolo Rosa; occurrenceID: 92FDD0A6-54DE-5166-B4DF-372F55037E84
- d. phylum: Arthropoda; class: Insecta; order: Hymenoptera; family: Chrysididae; genus: *Chrysis*; specificEpithet: *crossi*; country: Portugal; locality: Estoi; decimalLatitude: 37.1; decimalLongitude: -7.9; year: 2019; month: 5; day: 4; individualCount: 1; sex: M; lifeStage: Adult; recordedBy: P. Rosa & M. Jacobs; identifiedBy: Paolo Rosa; occurrenceID: 66AAAE02-1293-52EB-8267-4DBC53119F00
- e. phylum: Arthropoda; class: Insecta; order: Hymenoptera; family: Chrysididae; genus: *Chrysis*; specificEpithet: *crossi*; country: Portugal; municipality: Faro; locality: Montenegro; decimalLatitude: 37.0; decimalLongitude: -7.9; year: 2019; month: 5; day: 9; individualCount: 2; sex: M; lifeStage: Adult; recordedBy: P. Rosa & M. Jacobs (MJC); identifiedBy: Paolo Rosa; occurrenceID: 23B773FC-C44D-5C19-AE88-3CB26CCF3BA4
- f. phylum: Arthropoda; class: Insecta; order: Hymenoptera; family: Chrysididae; genus: *Chrysis*; specificEpithet: *crossi*; country: Portugal; locality: Rocha; decimalLatitude: 37.1; decimalLongitude: -8.5; year: 2021; month: 5; day: 26; individualCount: 6; sex: 5 M, 1 F;

lifeStage: Adult; recordedBy: M. Jacobs (MJC); identifiedBy: Paolo Rosa; occurrenceID: 4A481E5E-5CD0-5D06-A882-CA7D5B027CEC

Description

Female. Body length (holotype) 5.0 mm. Forewing length 3.5 mm.

Head. Vertex and frons with small, contiguous punctures (from 0.2× to 0.3× MOD) and polished interspaces below brow; transverse frontal carina faint; scapal basin medially transversally microridged, laterally with small punctures increasing diameter towards eye (Fig. 7D); malar spaces densely punctate, elongate (1.7× MOD), shorter than first flagellomere (2.0× MOD) and with short dense, silver setae; genal carina fully developed to mandibular insertion; clypeus mostly polished, sparsely punctate along anterior margin; clypeus elongate, subantennal area 1.7 MOD; medially notched and apically thickened. First flagellomere elongate, $l/w = 4$ (width taken at base of flagellomere). OOL 1.4× MOD; POL 2.0× MOD; MS 1.7× MOD; relative length of P:F1:F2:F3 = 1.0:1.6:0.9:0.7.

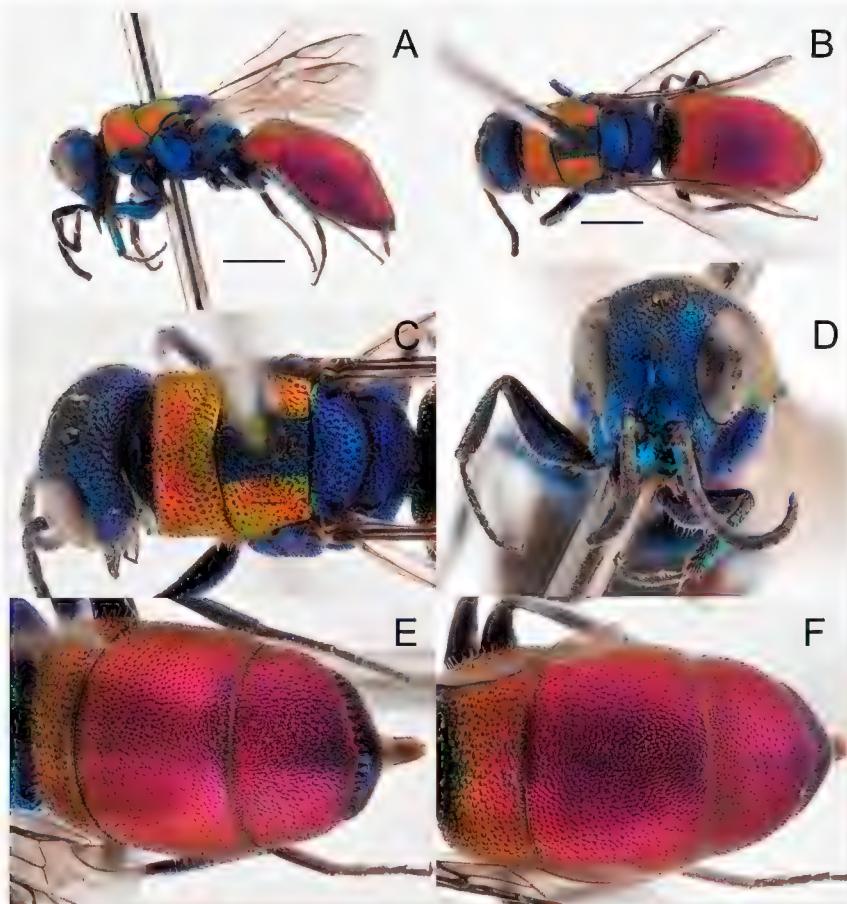


Figure 7. [doi](#).

Chrysis crossi Rosa, sp. nov., female, holotype. **A** habitus, lateral view; **B** habitus, dorsal view; **C** head and mesosoma, dorsal view; **D** head, frontal view; **E** metasoma, postero-lateral view; **F** metasoma, dorsal view. Scale bar: 1 mm.

Mesosoma. Medial pronotal line narrow and short, reaching half pronotal length; pronotum antero-laterally slightly bulging (Fig. 7); pronotal punctation double and interspaces polished with sparse minute dots; notaulus basally formed by small subrectangular foveae becoming smaller and rounded at apex; parapsidal signum as a linear depression; mesoscutellum dense puncture and irregular interspaces, antero-

medially corrugated and becoming polished towards base; scrobal sulcus of mesopleuron formed by large foveae aligned, limited to upper half; episternal sulcus formed by large and irregular, subsquare foveae; punctuation with dots on interspaces and larger punctures on mesepisterum; scutellar-metanotal suture deep and wide; metanotum with contiguous punctures, larger than other punctures on mesosoma; posterior propodeal projections slightly divergent; wing venation unmodified.

Metasoma. First tergum double punctate, with large punctures separated by small punctures on interspaces; second and third tergum double punctate, larger punctures smaller than those on first tergum; punctures on metasomal separated by polished interspaces (Fig. 7B, Fig. 7F); pit row composed by small, deep pits, apical margin of third tergum continuous, dark blue, medially arcuate; black spots of the second sternum large, covering almost all segment length, reaching median line.

Colouration. Head and mesosoma dark blue, pronotum and lateral areas of mesoscutum flame red, scutellum with light blue highlights; metasoma red to purplish, apical margin of third tergum blue. Scape, pedicel and first tergum black with weak greenish-metallic lustre, rest of flagellum black; tegula blue; metasomal venter black, with only a narrow blue thin line between the black spots and the apical margin of the second sternum. Legs blue, tarsi dark brown.

Male. Body length 5.0–6.0 mm. Similar to female in shape, sculpture and colouration. Malar space slightly shorter, scapal basin laterally covered by short, dense, appressed and silvery pubescence; blue segments of mesososoma with greenish reflection, propodeum and propodeal angles dorsally green to golden green; brown. Male genital capsule (Fig. 8A) with inner margin of the gonocoxa straight.

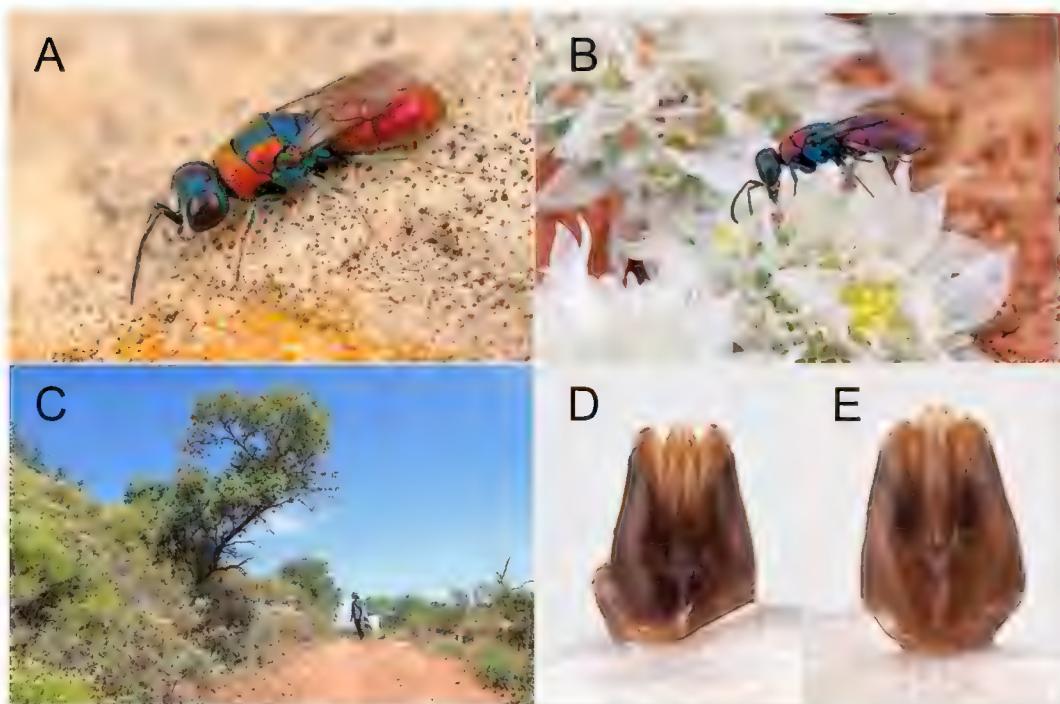


Figure 8. [doi](#)

Chrysis crossi Rosa, sp. nov. **A** male (Ph.: © M. Jacobs); **B** female (Ph.: © M. Jacobs); **C** collecting site of *Chrysis crossi* with Maarten Jacobs; **D** Male genital capsule; **E** Male genital capsule of *Chrysis phryne* from Italy (Emilia-Romagna, Oriano).

Diagnosis

Medium-sized, slender species (5–6 mm); head and mesosoma blue, pronotum and lateral areas of mesoscutum red; mesosoma dorsally red to purple, apical margin of third tergum blue; metasoma ventrally black, black spots on second sternum large, covering almost all surface and touching mid-line, without being clearly fused with each other; narrow stripe on apical margin of second tergum blue. Metasoma punctuation double, dense, with polished interspaces between the large and small punctures. *Chrysis crossi* sp. nov. is chromatically and morphologically similar to *C. phryne* Abeille de Perrin, 1878, but it is clearly separated genetically (see below). The main diagnostic characters to separate both sexes from *C. phryne* is the punctuation, which consists of distinct double punctures on the metasomal scutum, these being separated by polished interspaces (Fig. 7E, Fig. 9A), whereas in *C. phryne*, the punctuation is even and dense, without polished spaces. The metasomal venter is black in both sexes; black spots on second sternum large, covering almost all surface and touching mid-line with a narrow blue line between the black spots and the apical margin of the segment; in *C. phryne*, the sternum is clearly metallic green to golden green, with black spots distinctly separate from mid-line. The male genital capsule of the two species is different (Fig. 8) being narrower and more slender in *C. crossi* sp. nov., with the inner margin of the gonocoxa straight. Male can be also recognised by their brown tarsi, which are pale to yellowish in *C. phryne*.

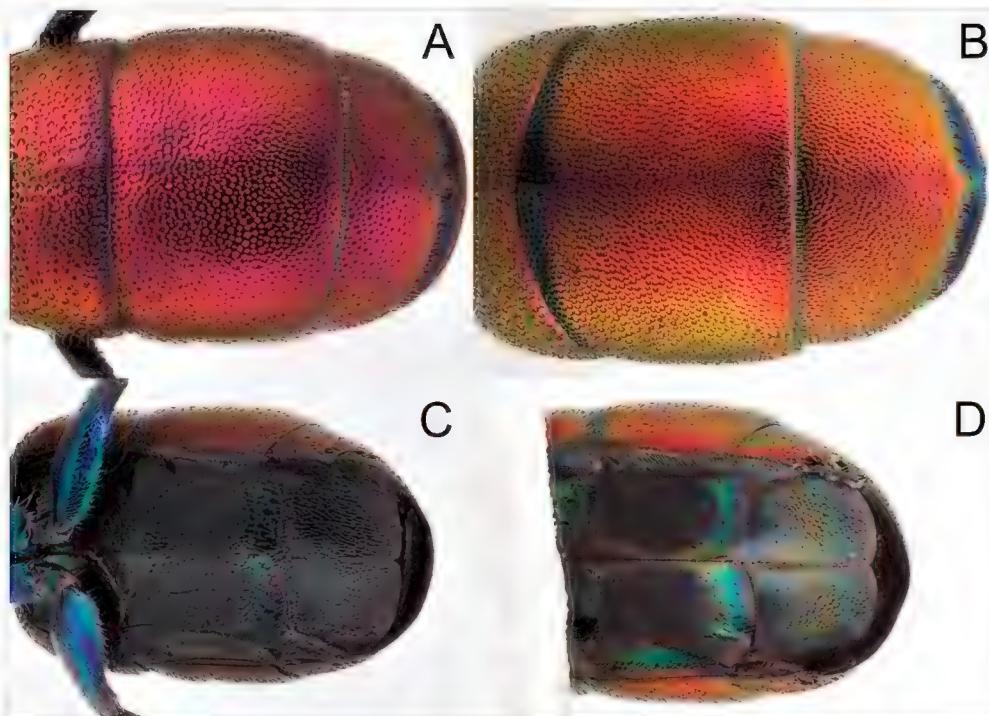


Figure 9. [doi](#)

A-B metasoma, dorsal view: **A** *Chrysis crossi* Rosa, sp. nov., male, paratype; **B** *Chrysis phryne* male, from Italy (Emilia-Romagna, Oriano); **C-D** metasoma, ventral view; **C** *Chrysis crossi* Rosa, sp., nov., male, paratype; **D** *Chrysis phryne* male, from Italy (Emilia-Romagna, Oriano).

Genetics: *Chrysis crossi* is very distinct genetically, being separated from an Italian specimen of *C. phryne* by 8.51%. There are no other specimens separated by less than a genetic distance of 10.0%.

Etymology

The specific epithet *crossi* (masculine) is dedicated to Ian Cross (Dorchester, Dorset, UK) for his active research on Portuguese Hymenoptera, including cuckoo wasps, many specimens of which were used for the current InBIO Barcoding Initiative work.

Distribution

Portugal (Algarve).

Ecology

Label information from Ian Cross reports that a male specimen was collected at an aggregation of *Melitturga caudata* Pérez, 1879 (Andrenidae), on the sand near empty snail shells. *Chrysis phryne* has been reported to attack *Osmia (Allosmia) melanura* Morawitz, 1871 (see Pauli et al. (2019)), but this cannot be the typical host across much of its range as, in Europe, *O. melanura* is restricted to southern Italy, North Macedonia and southern Ukraine (Müller 2022). It is likely that a different snail shell-nesting *O. (Allosmia)* is used instead, all known species of *O. (Allosmia)* using this nesting substrate. The likely host is *O. (Allosmia) rufohirta* Latreille, 1811 which is widely distributed across Europe and is the only *O. (Allosmia)* known from Portugal (Müller 2022), being common in the Algarve (Baldock et al. 2018). We suggest that *O. rufohirta* is the likely host of *C. crossi*, though this must be confirmed through direct observations.

Notes

According to Linsenmaier (1959), the *phryne* group includes only two species: *Chrysis circe* Mocsáry, 1889 and *C. phryne* Abeille de Perrin, 1878, with three subspecies *C. phryne* s.str., *C. phryne hebraeica* Linsenmaier, 1959 and *C. phryne burgenlandia* Linsenmaier, 1968. The types of these subspecies have been examined and *C. hebraeica* **stat. nov.** has to be considered to be a distinct species, based on morphological analyses, as it displays greater morphological differences from *C. phryne* s.str. than *C. crossi*. Comments on the specific status of *C. phryne burgenlandia* (known from Austria to Greece) should be postponed until genetic sequences are available, because the main diagnostic characters are based on body colouration only.

Finally, Mocsáry (1889) described *Chrysis destefanii*, based on the description of a specimen collected in Sicily by De Stefani-Perez and identified as *C. candens* by du Buysson (1888). The type of *Chrysis destefanii* is currently considered to be lost, as is large part of De Stefani's collection (Romano 2006). *Chrysis destefanii* was considered to be a synonym of *C. phryne* by Linsenmaier (1959) and Kimsey and Bohart (1991). Strumia and Yildirim (2009) identified a specimen from Turkey as *Chrysis destefanii*, yet this record may be related to *Chrysis hebraeica* or to an undescribed species of the similar *rubricata* group that has already been observed in the Middle East (PR, unpublished data). Based on the descriptions by du Buysson (1888) and Mocsáry

(1889) and, in particular, on the detail on the punctuation of the second tergum “régulière formée de points égaux, assez serrés”, the synonymy between *Chrysis destefanii* and *C. crossi* is excluded and De Stefani’s specimen would appear to be conspecific with *C. phryne*. Since the type of *Chrysis destefanii* is lost, we treat *C. destefani* as **nomen dubium**, until such a point that molecular data are available for Sicilian specimens.

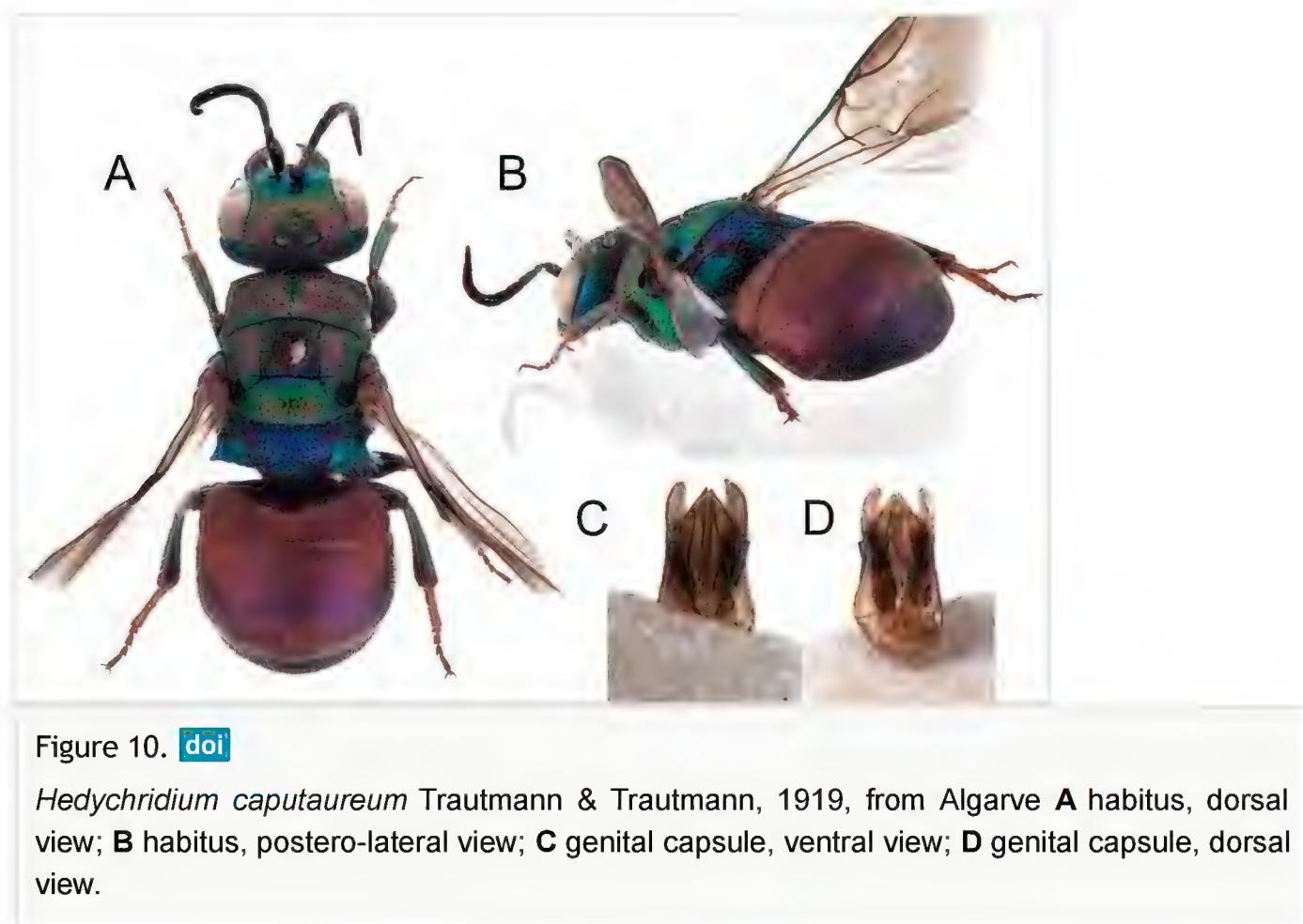
Analysis

Hedychridium caputaureum Trautmann & Trautmann, 1919 and *Hedychridium chloropygum* du Buysson, 1888

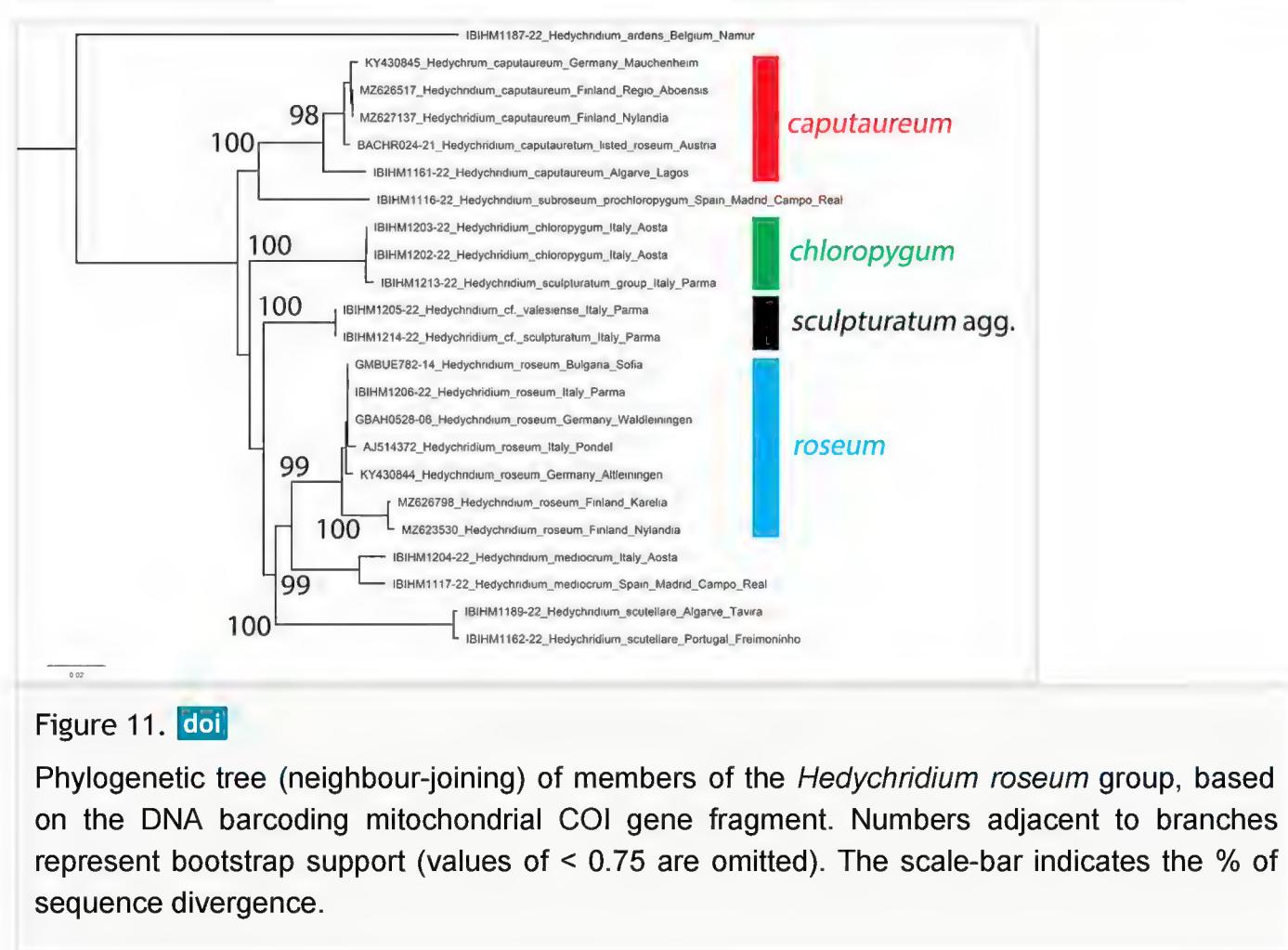
Arens (2010) considered *Hedychridium caputaureum* Trautmann & Trautmann, 1919 to be a subspecies of *H. chloropygum* du Buysson, 1888, based on morphological affinities and noticeable variation in surface sculpture. According to Arens (2010), *H. chloropygum* s.str. is limited to south-western Europe, whereas *H. chloropygum caputaureum* is distributed in northern, central and south-eastern Europe. The main difference between these two taxa is based on the colouration. In *H. chloropygum* s.str., the anterior part of the body is richly adorned with copper or gold and the metasoma is metallic blue to violet on the third tergum, to a varying extent and intensity. In *H. chloropygum caputaureum*, the anterior part of the body may have a similar colouration, although Nordic and central European specimens may be darker, with faint coppery or golden reflections (described as *H. chloropygum densum* Linsenmaier, 1959 and synonymised by Arens (2010) with *H. chloropygum caputaureum*); however, the metasoma is always without metallic reflections. The two taxa can additionally be separated by the denser and coarser punctuation of the metasoma in *H. caputaureum* which is apparently locally variable (Arens 2010).

The specimen IBIHM1161-22 collected in the Algarve (Lagos, Fig. 10) shows a rich golden colouration of the anterior part of the body and third tergum is entirely violet, with the metasoma densely punctate. DNA barcodes demonstrate there is moderate genetic differentiation between the Portuguese specimen and *H. caputaureum* from northern and central Europe (Fig. 11), being separated by an average of 2.50% (range 2.43-2.59%). The clade of *H. caputaureum* from Austria, Finland and Germany shows low average intraspecific distance of 0.19% (range 0.00-0.34%) and has bootstrap support of 98%.

However, the Portuguese specimen is much more strongly separated from two sequences of *H. chloropygum* from Italy, showing average genetic differentiation of 8.43% (range 8.36-8.51%). When including the Portuguese specimen within *H. caputaureum*, the two clades are separated by an average interspecific genetic distance of 8.56% (range 8.36-9.00%). We, therefore, consider *H. caputaureum* and *H. chloropygum* to be two different species and include Portuguese material within *H. caputaureum*, with the observed genetic distance considered to be variation, given the geographic distance between southern Portugal and Germany. The overall distribution of *H. caputaureum* must be revised, as this Portuguese specimen is the first reported record of this taxon in south-western Europe. Additional genetic samples from Spain and France are likely to fall between the Portuguese and central/northern European sequences.

Figure 10. [doi](#)

Hedychridium caputaureum Trautmann & Trautmann, 1919, from Algarve **A** habitus, dorsal view; **B** habitus, postero-lateral view; **C** genital capsule, ventral view; **D** genital capsule, dorsal view.

Figure 11. [doi](#)

Phylogenetic tree (neighbour-joining) of members of the *Hedychridium roseum* group, based on the DNA barcoding mitochondrial COI gene fragment. Numbers adjacent to branches represent bootstrap support (values of < 0.75 are omitted). The scale-bar indicates the % of sequence divergence.

The subspecies *H. chloropygum berberiacum* Linsenmaier, 1959 from Algeria and Morocco shows a similar colouration to the Algarve specimen, though it has more extensively metallic violet colouration laterally on the metasoma, but also sparser punctuation (Rosa et al. 2022). Genetic analyses are needed to clarify the placement of this taxon, but for the

moment, we consider it to be the northern African subspecies of *H. chloropygum*, based on its shallow and sparse punctuation.

***Hedychrum rutilans* Dahlbom, 1854 and *Hedychrum viridiaureum* Tournier, 1877 sp. resurr.**

Hedychrum rutilans Dahlbom, 1854 is one of the most common European cuckoo wasp species, known to be a cleptoparasite of *Philanthus* species (Lisenmaier 1997a). In addition to typical cleptoparasitic behaviour, the female does not have to enter the host nest for ovipositing, but can oviposit directly on the prey (*Apis mellifera* Linnaeus) while it is being transported to the nest by the host (Veenendaal 1987, Baumgarten 1995). This species is also known in literature as *H. intermedium* sensu auctorum for an incorrect interpretation of the type materials (Rosa and Xu 2015). Lisenmaier (Lisenmaier 1959, Lisenmaier 1997a, Lisenmaier 1997b) considered three European subspecies, namely: *rutilans* s.str., ssp. *viridiaureum* Tournier, 1877 and ssp. *viridauratum* Mocsáry, 1889.

Two specimens from the same sampling locality in central Spain (Segovia, Bernuy de Porreros, IBIHM1120-22 and IBIHM1121-22) were both identified as *H. rutilans*, but are separated by a genetic distance of 5.27%. Integrating all newly-acquired sequences and sequences from BOLD and GenBank (some without identifications beyond *Hedychrum* sp.) shows that *H. rutilans* s.l. comprises two taxa (Fig. 12). Sequences from Spain, Italy, Germany, Finland and Bulgaria belong to *H. rutilans* s. str. (Fig. 12). They show low average intraspecific distance of 0.55% (range 0.00-1.48) and form a clade with bootstrap support of 100. Sequences from Portugal, Spain and western Germany (Rhineland-Palatinate) also show a low average intraspecific distance of 0.16% (range 0.00-0.33) and have bootstrap support of 100%. The two clades are separated by an average genetic distance of 5.16% (range 4.78-5.77%).

Following Lisenmaier (1997a), the second clade is called *Hedychrum viridiaureum* Tournier, 1877 sp. resurr. Lisenmaier (Lisenmaier 1959, Lisenmaier 1997a) noted differences between these two taxa and employed a subspecific framework. *Hedychrum rutilans* s. str. (Fig. 13) is usually larger and Lisenmaier noted a host association with *Philanthus coronatus* (Thunberg, 1784), whereas *H. viridiaureum* (Fig. 14) is usually smaller and is associated with the respectively smaller host *Philanthus triangulum* (Fabricius, 1775). Based on the DNA barcodes presented here, combined with the distributional framework of Lisenmaier, in Europe, *H. rutilans* s. str. appears to be more widely distributed, from central Iberia across the continent, whereas *H. viridiaureum* appears to be restricted to Iberia and western Europe, to Switzerland, western Germany and north to Belgium and the Netherlands. Detailed revision is necessary to clarify these range limits and all host associations as, in some regions where *H. rutilans* s.str. occurs, only *P. triangulum* is present. Our observations would support the host associations noted by Lisenmaier, with additional points. Very small individuals of *H. viridiaureum* can be found in Iberia (such as IBIHM-1183-22) where they are associated with *Philanthus pulchellus* Spinola, 1842 that is smaller than *P. triangulum*. Equally, in central Spain, the large-bodied *Philanthus dufourii* Lucas, 1849 is much more frequently encountered than *P.*

coronatus and, hence, this is likely the principal host of *H. rutilans* s. str. in this region. The correct placement of *H. rutilans viridiauratum* Mocsáry, 1889, described from Algeria (types examined) and cited by Linsenmaier (Linsenmaier 1959, Linsenmaier 1997a) from the Iberian Peninsula is unclear and must be evaluated by means of molecular analyses. It could be a synonym of *H. viridiaureum* Tournier or a separate northern African species.

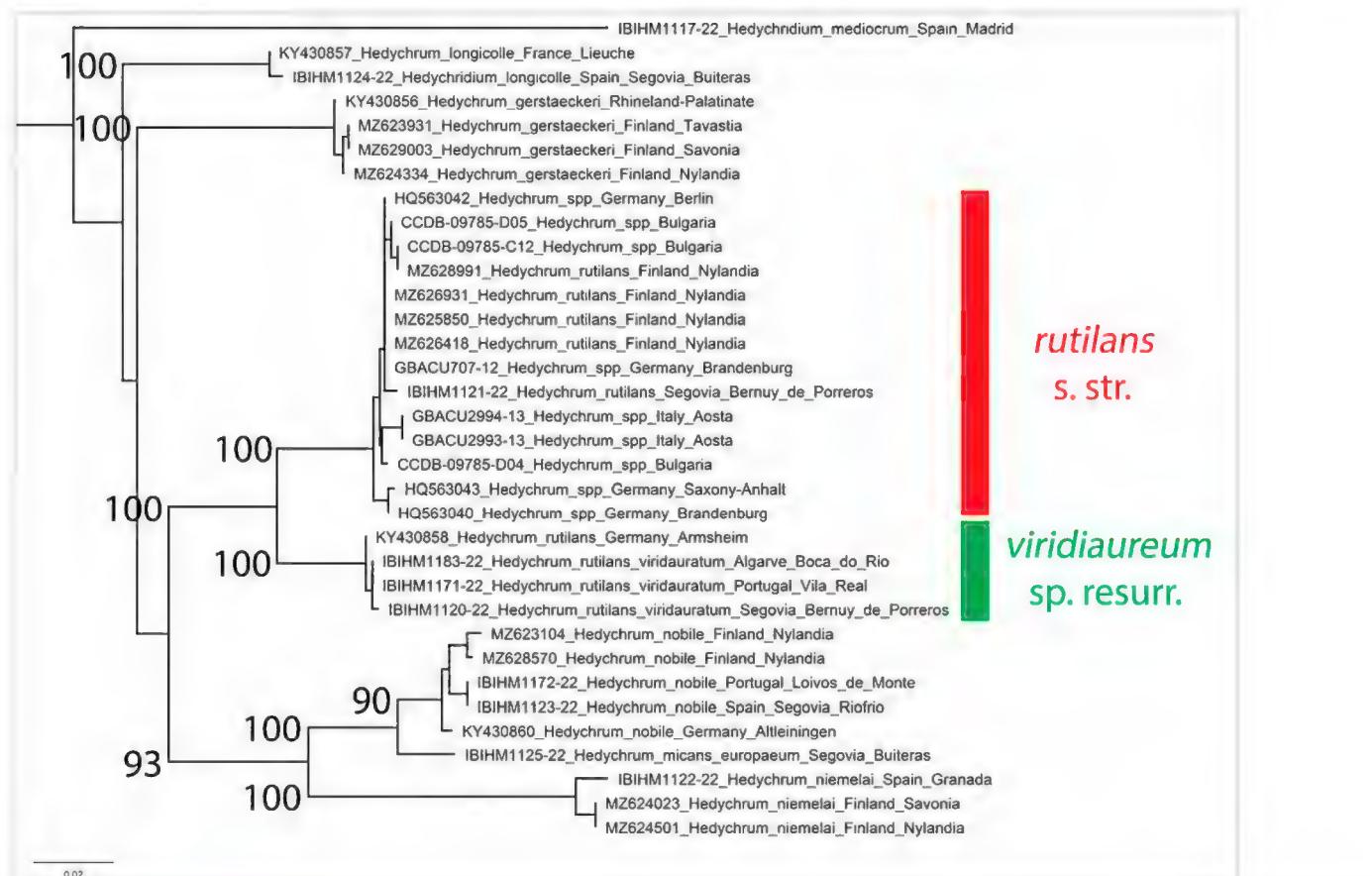


Figure 12. [doi](#)

Phylogenetic tree (neighbour-joining) of *Hedychrum* species, based on the DNA barcoding mitochondrial COI gene fragment. Numbers adjacent to branches represent bootstrap support (values of < 0.75 are omitted). The scale-bar indicates the % of sequence divergence.

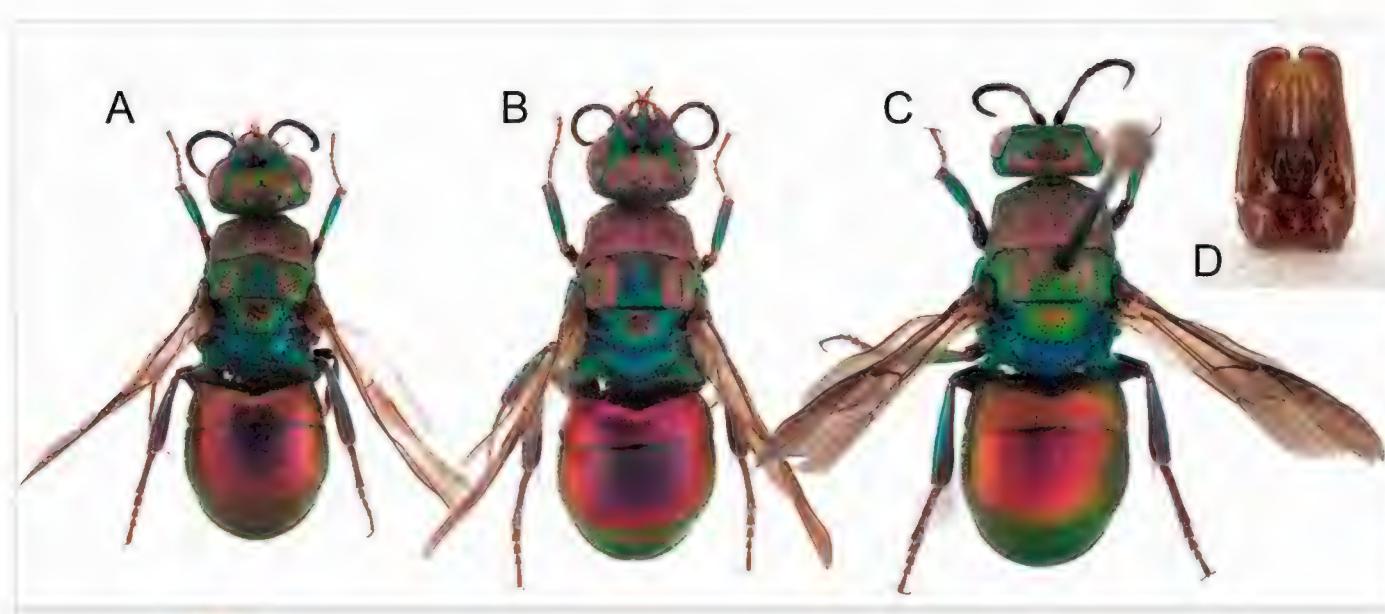
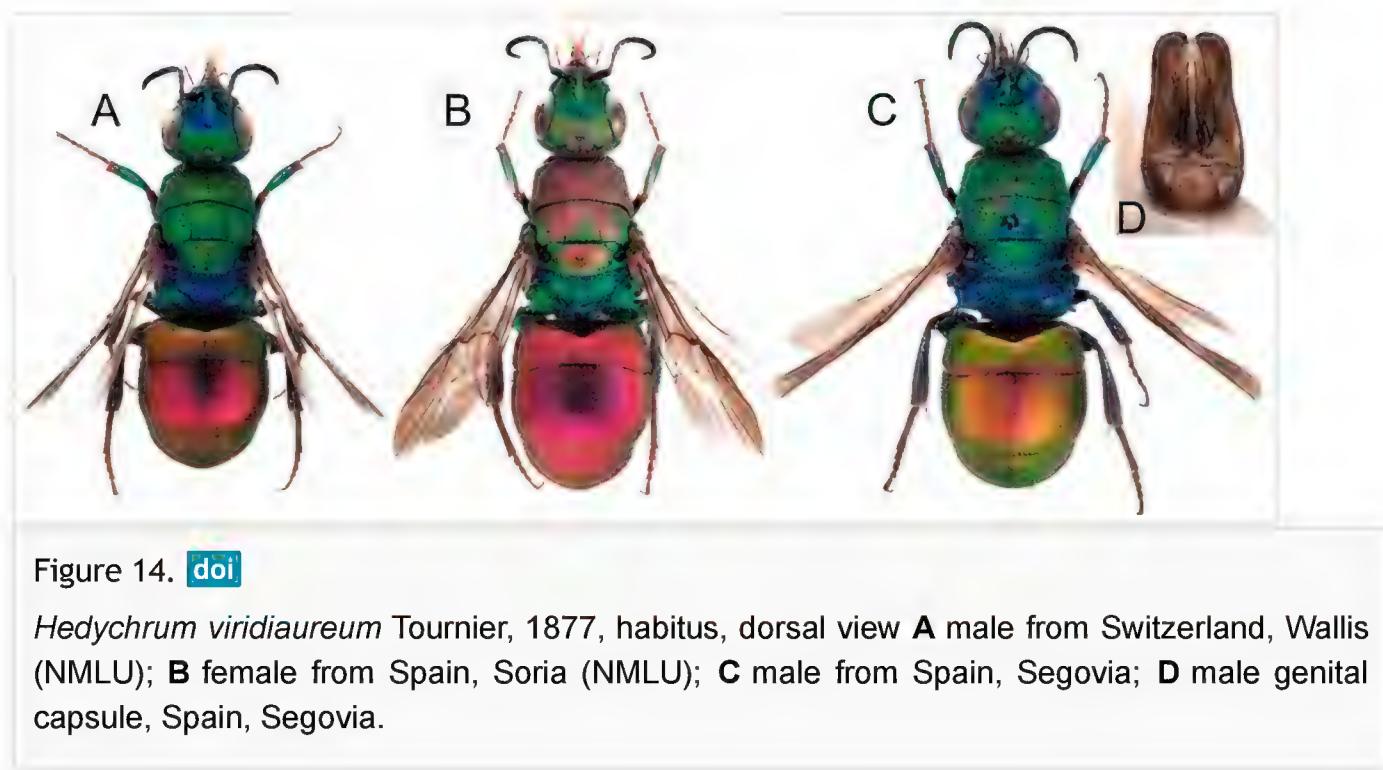


Figure 13. [doi](#)

Hedychrum rutilans Dahlbom, 1854, habitus, dorsal view **A** male from Austria, Hainburg (NMLU); **B** female from Austria, Hainburg (NMLU); **C** female from Spain, Segovia; **D** male genital capsule, Austria (NMLU).



Philoctetes punctulatus (Dahlbom, 1845) and *Philoctetes parvulus* (Dahlbom, 1845)

Philoctetes parvulus (Dahlbom, 1845) was considered to be a valid species by Rosa and Soon (2012) following type examination. Based on the consistently smaller size, dark colouration and different punctuation of both sexes in comparison with *P. punctulatus* (Fig. 15), specimens of *P. parvulus* were considered to be distinct. However, DNA barcodes (Fig. 16) demonstrate that the small individuals displaying the typical morphology of *P. parvulus* (INV12750, INV12749) show almost no genetic differentiation from *P. punctulatus* (0.00% and 0.02%). The differences in morphology are, therefore, not considered to be species-specific and are probably caused or exaggerated by the smaller body size. *Philoctetes parvulus* is, therefore, considered to be a synonym of *P. punctulatus*.

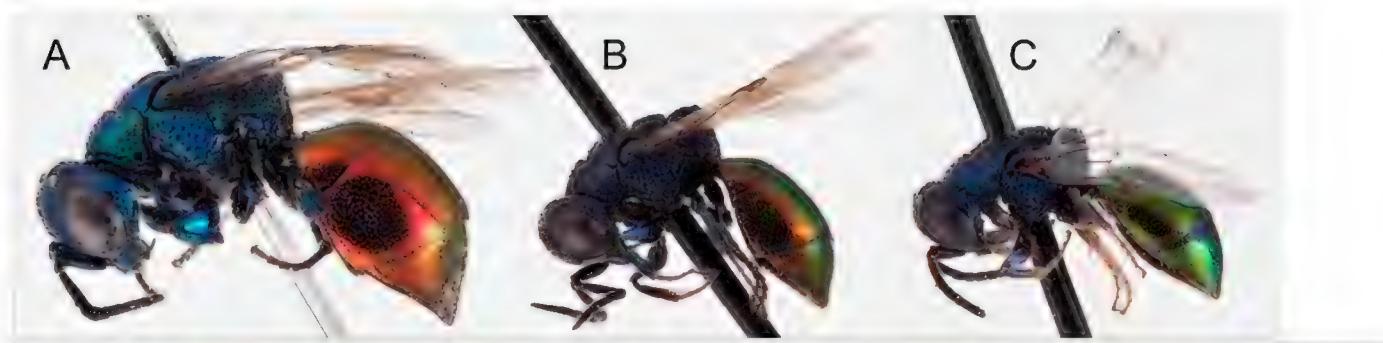


Figure 15. [doi](#)
Philoctetes punctulatus (Dahlbom, 1854), habitus, lateral view **A** female from Spain, Segovia; **B** female from Portugal, Tavira; **C** male from Portugal, Tavira.

Chrysis lusitanica Bischoff, 1910

The identity of *Chrysis lusitanica* Bischoff, 1910 (Fig. 17) has to date remained unclear. Kimsey and Bohart (1991) synonymised *Chrysis sculpturata* Mocsáry, 1912 with *C.*

lusitanica, but the examination of both types revealed the occurrence of two distinct species (Rosa et al. 2017, PR, unpublished data). Soon et al. (2014) DNA barcoded and revalidated *C. sculpturata* in the clade of *C. ignita* (Linnaeus, 1758), whereas the placement of *C. lusitanica* remained unknown.

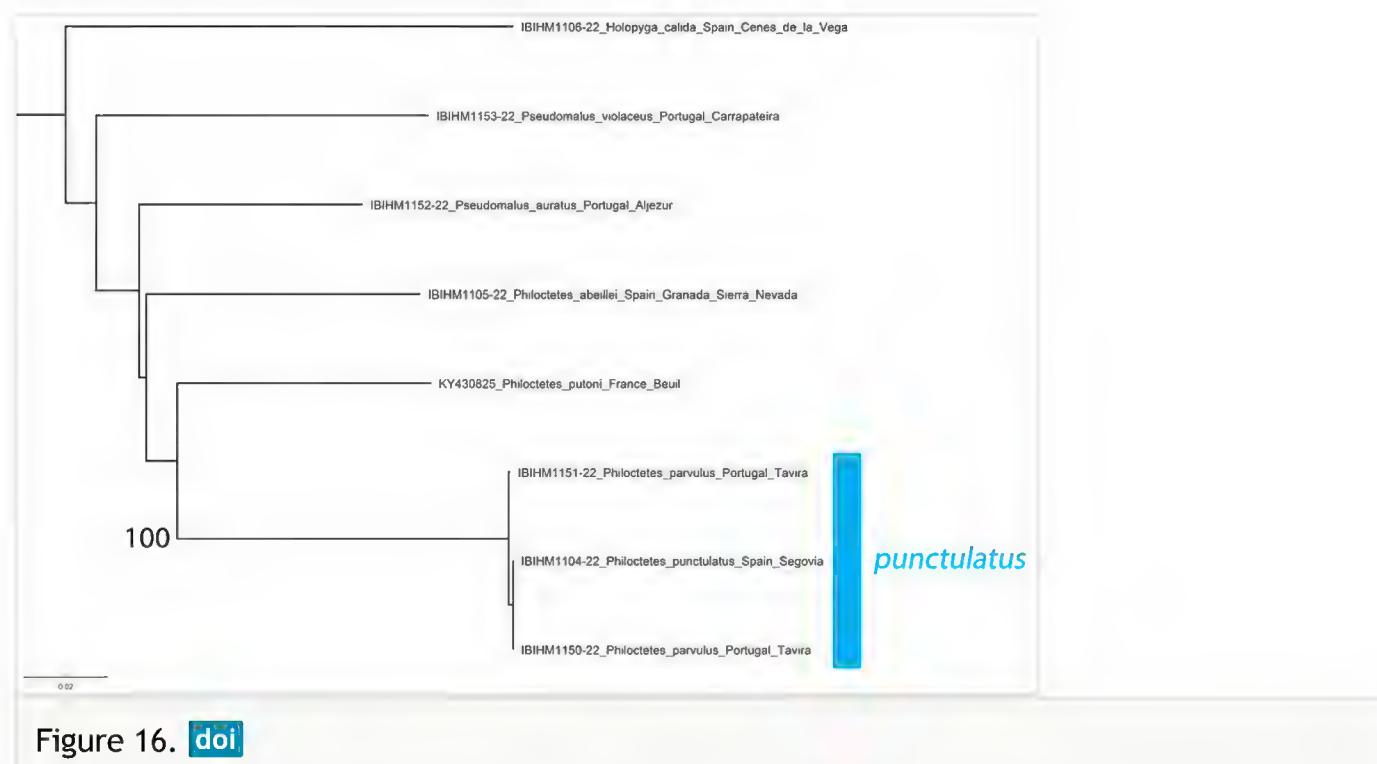


Figure 16. [doi](#)

Phylogenetic tree (neighbour-joining) of Elampini species with a focus on *Phiocetes*, based on the DNA barcoding mitochondrial COI gene fragment. Numbers adjacent to branches represent bootstrap support (values of < 0.75 are omitted). The scale-bar indicates the % of sequence divergence.

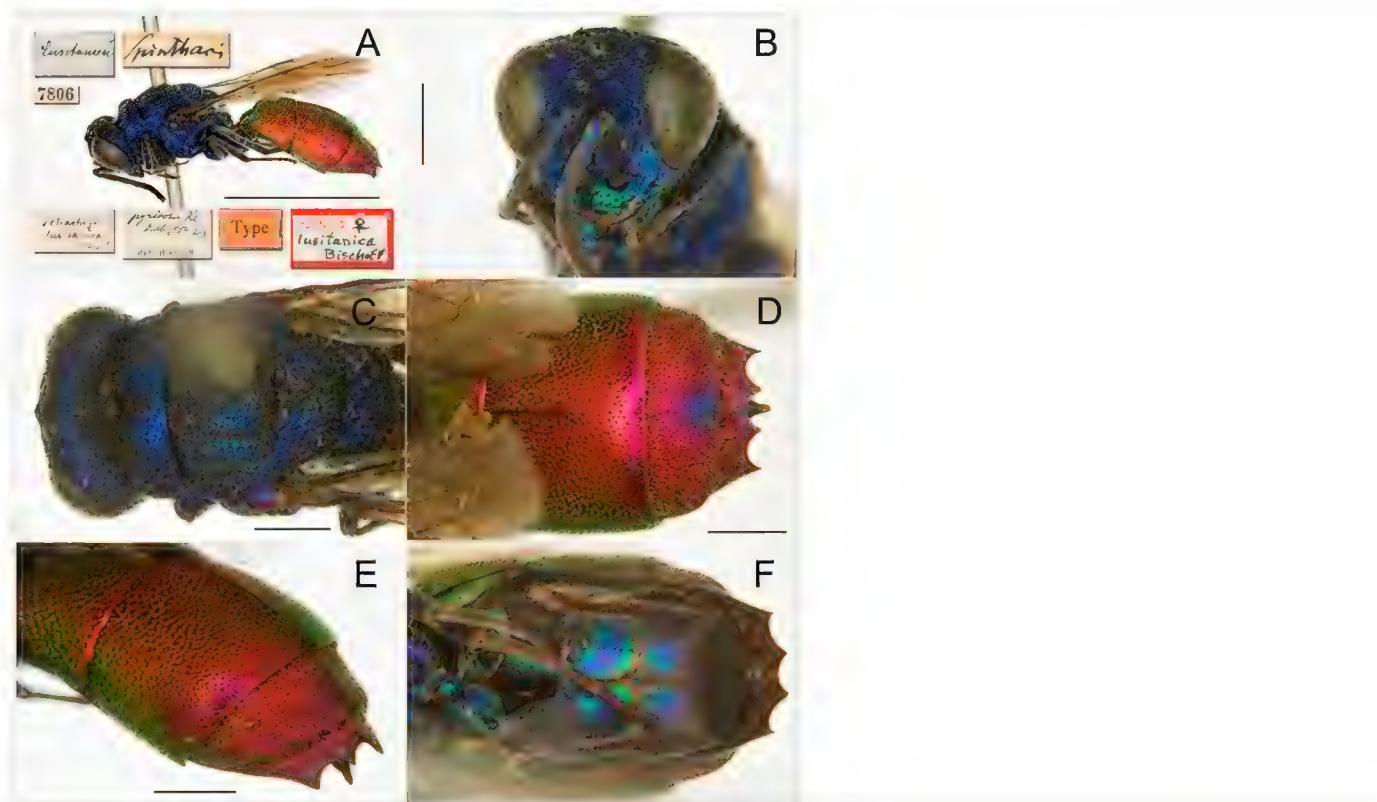
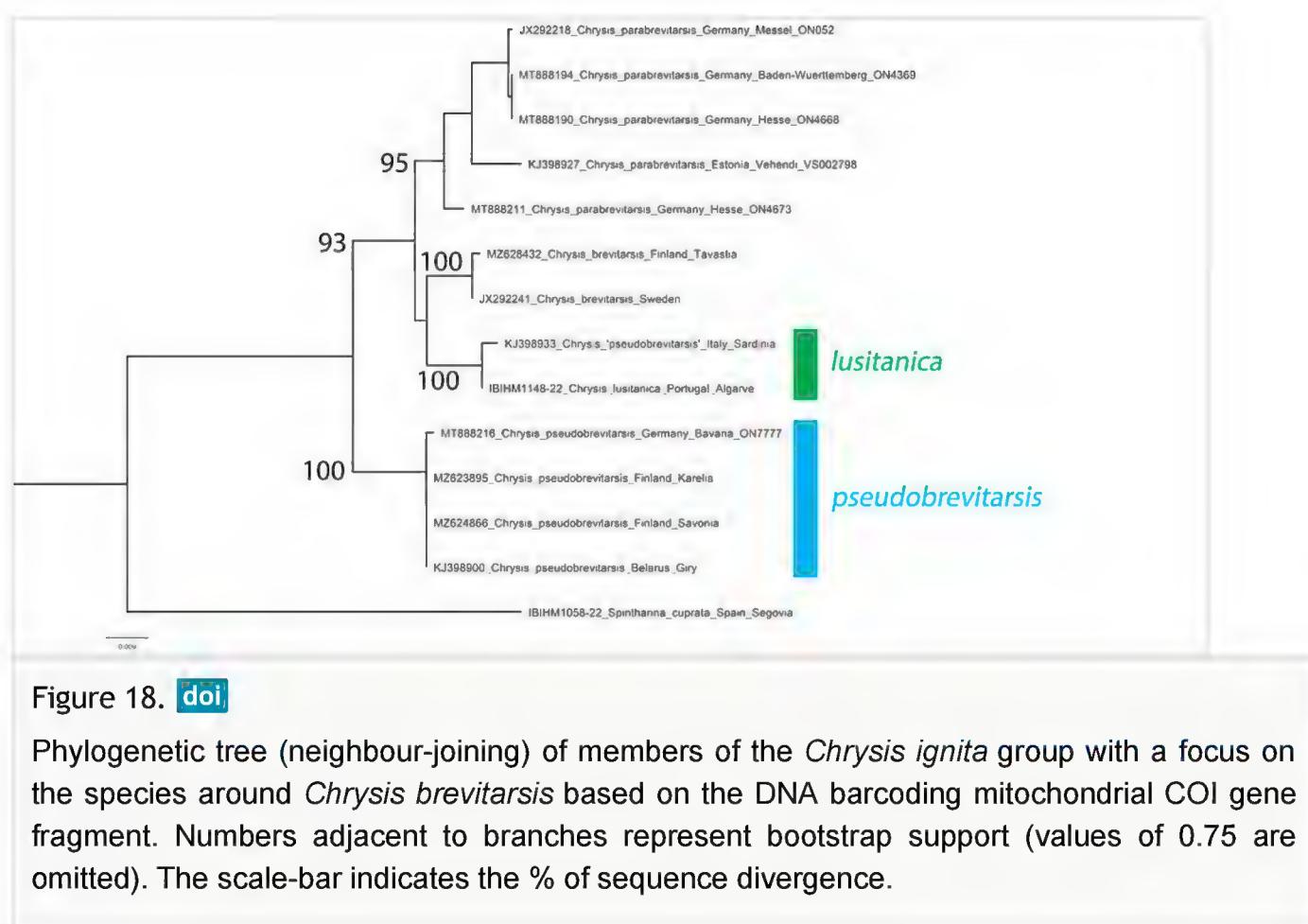


Figure 17. [doi](#)

Chrysis lusitanica Bischoff, 1910, female, holotype (MfN) **A** habitus, lateral view; **B** head, frontal view; **C** head and mesosoma, dorsal view; **D** metasoma, dorsal view; **E** metasoma, postero-lateral view; **F** metasoma, ventral view.

The DNA barcode of a recently-collected specimen places *C. lusitanica* in the clade of *C. brevitarsis* Thomson, 1870, a well-studied group after the molecular works of Soon et al. (2014) and Orlovskyté et al. (2016). In the framework of these previous projects, we can conclude that *C. lusitanica* is a member of the *brevitarsis* clade and that it is also present in Sardinia (first record for Italy, previously published as *C. pseudobrevitarsis* Linsenmaier, 1951 by Soon et al. (2014)). *Chrysis lusitanica* can be easily separated from the other species of this clade by the small, even and dense punctures on the second tergum, these punctures being smaller or similar to those on the first tergum and the mesosoma densely punctate, with uniform dark blue colouration.

Chrysis lusitanica (including the Sardinian specimen that differs from the Portuguese specimen by 0.46%) is strongly separated from *C. pseudobrevitarsis* by an average interspecific distance of 4.07% (range 3.80-4.41%, Fig. 18). It is less strongly separated from *C. brevitarsis* by an average interspecific distance of 2.28% (range 2.05-2.43%), but because intraspecific variation is low (0.46% and 0.16%, respectively), both the *C. lusitanica* and *C. brevitarsis* clades have bootstrap support of 100%. Genetic differentiation within the *brevitarsis* clade is generally low, with *C. brevitarsis* separated from *C. parabrevitarsis* by an average interspecific distance of 3.06% (range 2.46-3.50%). In this context, *C. lusitanica* is considered to be a consistently differentiated species.



Chrysis scutellaris marteni Linsenmaier, 1951

Linsenmaier (1951) described a Spanish subspecies of *Chrysis scutellaris* Fabricius, 1794, based on its larger size (9.0-10.5 mm) and stocky aspect, with the apical margin of the third tergum indistinctly undulate. This form can be collected in sympatry with the

nominotypical species. The two specimens DNA barcoded here conform morphologically to *C. scutellaris marteni* and show a genetic distance of 6.26% and 6.42% from a sample of *C. scutellaris* from Eisenberg (Rhineland-Palatinate) in Germany ([KY430717](#)). The subspecies may, therefore, be distinct, but additional samples are required, particularly of typical *C. scutellaris* from Iberia.

***Chrysis splendidula* group**

The *splendidula* group currently includes twelve Palearctic species, yet the real number of the species in this group is unclear and requires detailed revision. Several morphospecies are found in the Mediterranean region, in particular those related to the subgroup of *C. rutilans* Olivier, 1790 and identified with the name *Chrysis insperata* Chevrier, 1870, including small and slender species. Other morphospecies closely related to *C. rutilans* are awaiting description (PR, unpublished data). The identity of specimen IBIHM1090-22 from the Sierra Nevada, tentatively identified as *C. rutilans*, is unclear, but it may represent another undescribed taxon within this group as it is clearly separated from *C. rutilans* sequences from Finland.

Within the *splendidula* group, we identify *Chrysis maroccana* Mocsáry, 1883 as a species closely related to *C. splendidula* Rossi, 1790, which lacks the raised apical margin at the apex of the second tergum. This species was previously reported from Morocco, Sardinia and Corsica (Lisenmaier 1987). Mingo (1994) recorded *C. maroccana* from Portugal and Spain, but her identifications clearly refer to another species already known from Iberia which was not mentioned in the monograph, namely *C. continentalis* Lisenmaier, 1959. Specimen IBIHM1141-22 from southern Portugal (Algarve, Praia do Barril) is strongly separated from two sequences of *C. splendidula* from Italy by an average of 7.67% (range 7.49-7.84%, Fig. 19). We suspect that the Portuguese specimen represents *C. maroccana*, but additional sampling and genetic sequences from Morocco are required for confident determination, given the complexity within this species group

***Stilbum westermanni* Dahlbom, 1845 sp. resurr.**

Lisenmaier (1959) listed six subspecies of *Stilbum calens* (Fabricius, 1781) distributed from Europe to China and he separated the two subspecies *S. calens zimmermanni* Lisenmaier, 1959 and *S. calens subcalens* Lisenmaier, 1959 (nec Mader 1933, an unavailable name) (Lisenmaier 1959, Lisenmaier 1997a, Lisenmaier 1997b). This latter taxon was later identified as *S. calens wesmaeli* Dahlbom, 1845 (Lisenmaier 1997a: 134, Lisenmaier 1997b: 287, Lisenmaier 1999: 254). However, Rosa and Vårdal (2015) discovered that the type of *Stilbum wesmaeli* is actually related to *S. cyanurum* (Forster, 1771) and the first available name for this taxon is *Stilbum westermanni* Dahlbom, 1845. Rosa and Vårdal (2015) considered this to be a subspecies of *Stilbum calens*, following Lisenmaier's subspecific interpretation (Lisenmaier 1959, Lisenmaier 1997a, Lisenmaier 1997b).

Genetic results unambiguously support a species-level difference between *S. calens westermanni* from Spain and Portugal and *S. calens zimmermanni* from Italy (Fig. 20). These taxa are separated by an average genetic distance of 6.62%, with bootstrap support of 100% for each clade. Both taxa are well-separated from *S. cyanurum* (Fig. 21A), by an average of 6.68% for *S. calens westermanni* and by 6.86% for *S. calens zimmermanni*. Though collected over a large area from Portugal to South Africa, *S. cyanurum* shows low intraspecific variability, with average separation of 1.26% (range 0.47-1.89%). *Stilbum westermanni* stat. nov. is, therefore, restored to species status (Fig. 21B). For now, we follow the interpretation of Lisenmaier that material from Central Europe should be referred to as *S. calens zimmermanni* until genetic samples are available from Siberia, the *locus typicus* of *S. calens* s. str.

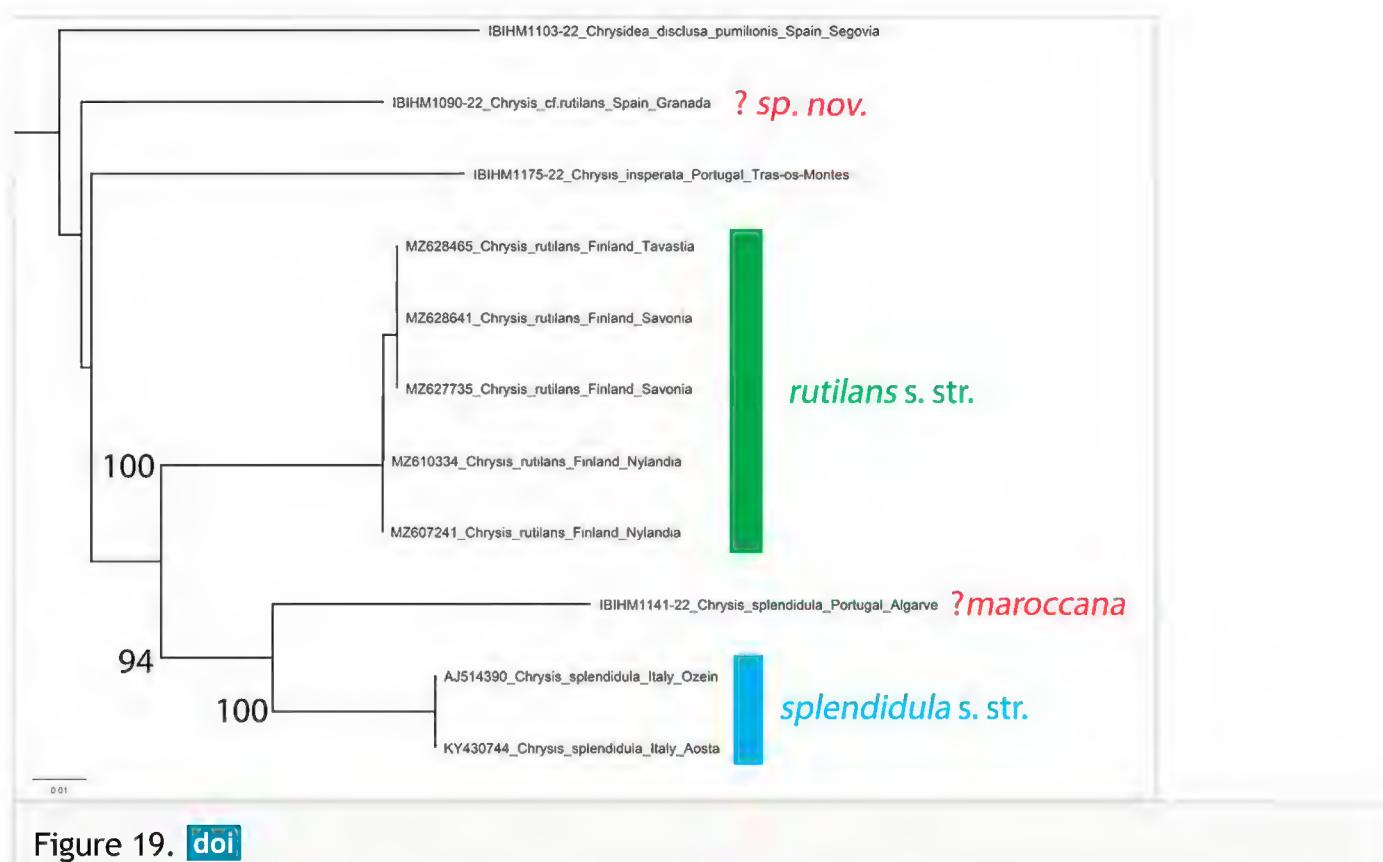
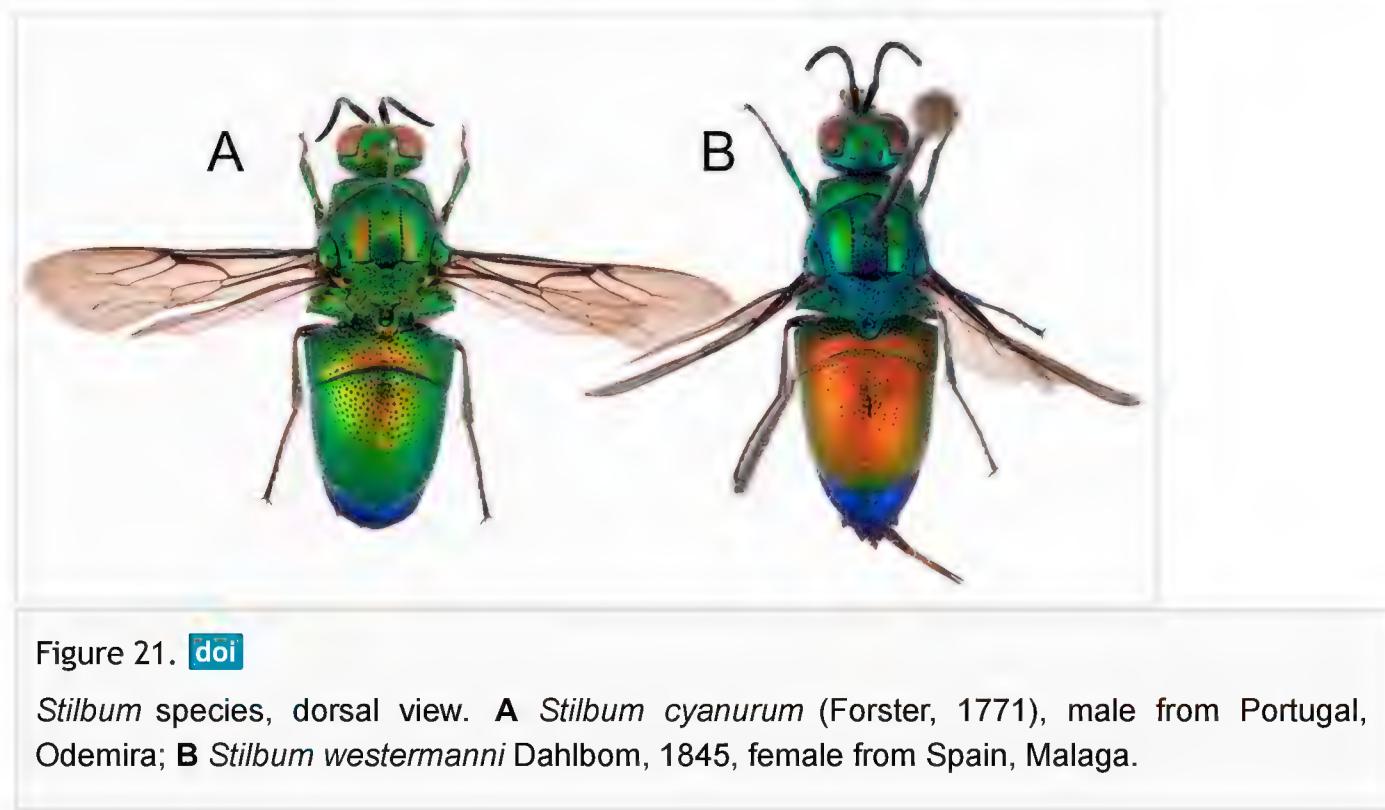
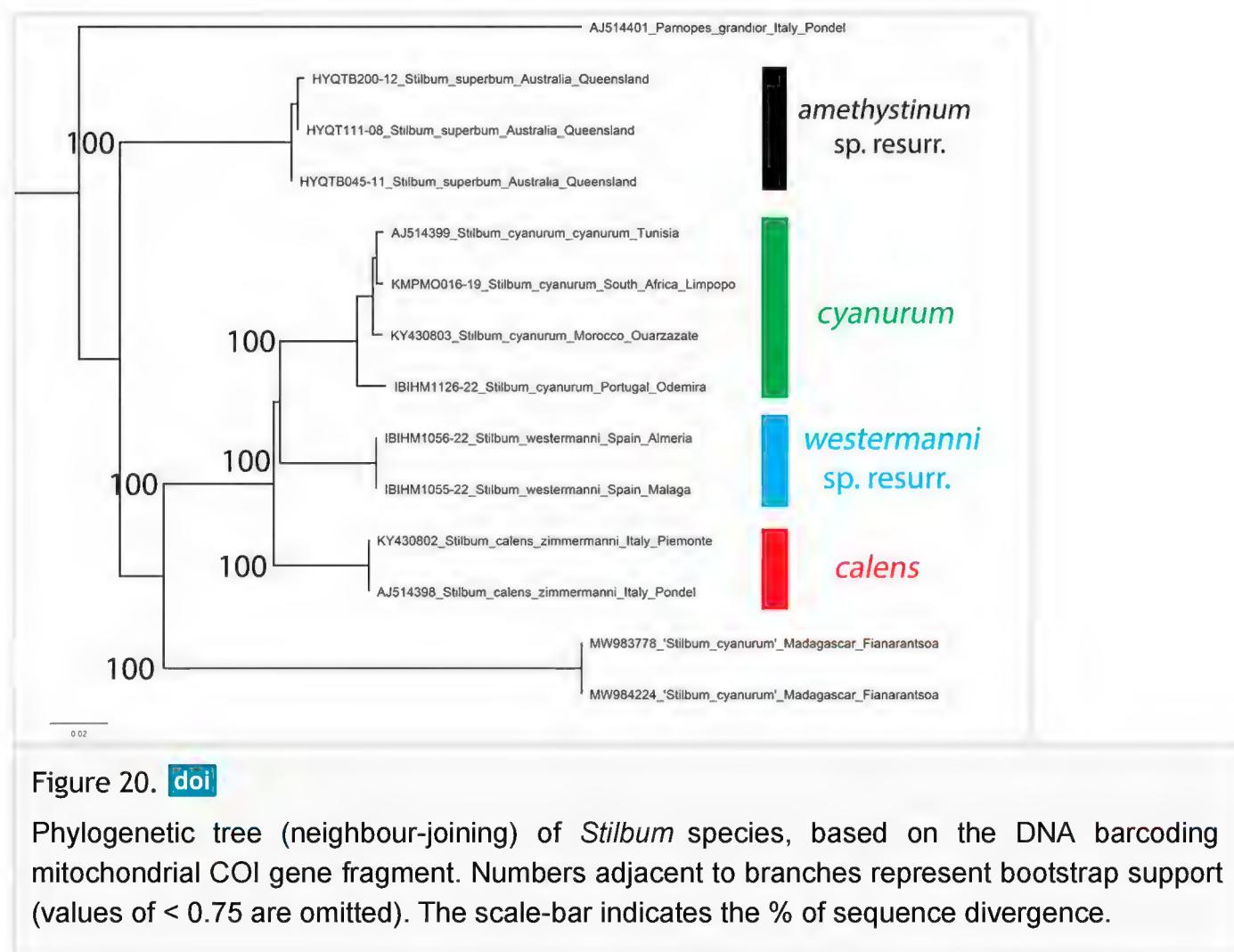


Figure 19. [doi](#)

Phylogenetic tree (neighbour-joining) of the *Chrysis splendidula* group, based on the DNA barcoding mitochondrial COI gene fragment. Numbers adjacent to branches represent bootstrap support (values of < 0.75 are omitted). The scale-bar indicates the % of sequence divergence.

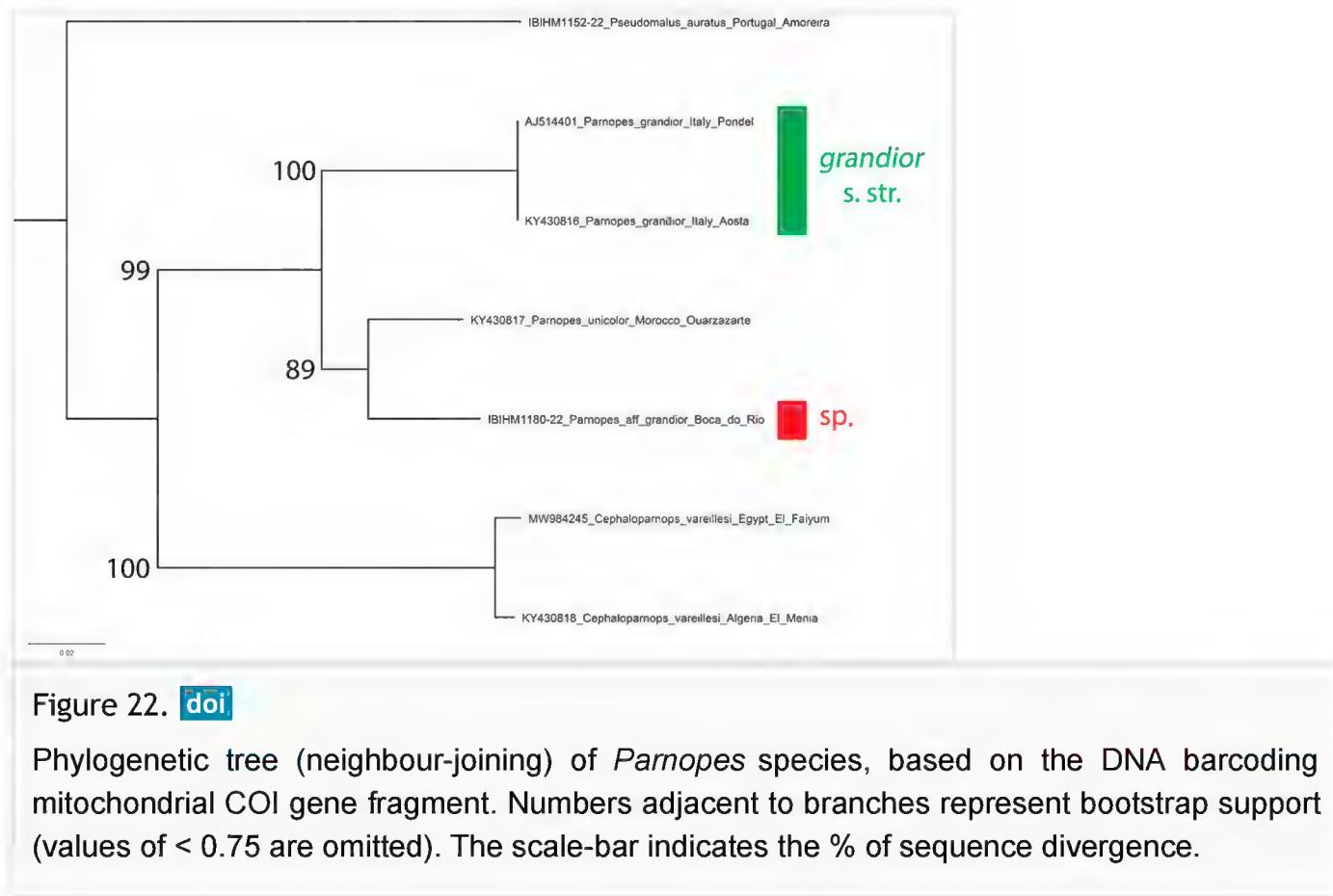
The specimens DNA barcoded and identified in BOLD as *Stilbum cyanurum* from Madagascar ([MW983778](#) and [MW983223](#)) are clearly distinct and actually belong to the species *Stilbum viride* Guérin-Méneville, 1842, the sole and endemic Madagascan *Stilbum* (Kimsey and Bohart 1991). Additionally, samples from Australia identified as 'S. superbum' are also clearly distinct. However, the name 'Stilbum superbum' is unavailable and it is likely an incorrect spelling of *Stilbum splendidum* (Fabricius, 1775) that has recently been used on online sites. The Australian *Stilbum* species is clearly morphologically different from all other known species, but its taxonomic status has been confused. In literature, it has been commonly referred to as *S. splendidum* auct. or *S. amethystinum* auct. The type of the first taxon proved to be morphologically conspecific with *S. cyanurum* and considered to be a subjective synonym by Kimsey and Bohart (1991). The second was

also considered to be a subjective synonym of *S. cyanurum* by Kimsey and Bohart (1991), but two syntopic specimens in London (Banks Collection, Natural History Museum) belong to a morphologically separated species, characterised by smaller size and short malar spaces. Since this taxon is morphologically and genetically distinct, we here resurrect *S. amethystinum* (Fabricius, 1775) sp. resurr. from its previous synonymy with *S. cyanurum*. We also designate here the lectotype of *Chrysis amethystina* Fabricius, 1775 with one of the two specimens housed in the Banks Collection.



Parnopes sp.

The genetic sequence of this *Parnopes* specimen is strongly separated from the sequence of *P. grandior* from Italy by an average of 8.71% (Fig. 22). It is closer to a *P. unicolor* sequence from Morocco, but is still separated by 5.18%. Taken together, this taxon and *Parnopes unicolor* form a clade with bootstrap support of 89%, strongly separated from the *P. grandior* clade that has bootstrap support of 100%.



The discovery of another *Parnopes* species in the Iberian Peninsula is not so surprising as it seems, even though a name currently cannot be confidently assigned to this taxon. In recent years, a new species from Sardinia, *Parnopes linsenmaieri* Agnoli, 1995 (described as subspecies of *Parnopes grandior*) was described and another species was found through DNA barcoding Bulgarian specimens (BOLD, unpublished sequences). However, several new species of West Palearctic *Parnopes* will be described in an upcoming revision. These species have been overlooked because, classically, only three species were considered to be valid in the West Palearctic: *Parnopes grandior* (known from Europe to central Asia), *P. unicolor* (northern Africa) and *P. glasunowi* (western Asia to central Asia) and specimens were identified, in part, based on the collecting locality and, in part, on body colouration. In this sense, all the variations and subspecies of *P. grandior* were considered to be only colour variation (Kimsey and Bohart 1991). The Portuguese female barcoded may be related to *Parnopes marokkanus* Trautmann, 1927, a taxon not mentioned by Kimsey and Bohart (1991), Linsenmaier (1959), Linsenmaier (1968), Linsenmaier (1997b) and Linsenmaier (1999). In any case, many more genetic sequences and analyses are needed to understand the limits of variability within this genus. In fact, colouration is still seemingly very variable within populations, but could also represent the presence of valid sibling species.

New additions to the Portuguese fauna

Thanks to this barcoding project, we analysed and added for the first time the following taxa to the list of the Portuguese species:

Hedychridium caputaureum Trautmann & Trautmann, 1919

Hedychridium cupritibiale Linsenmaier, 1987

Hedychridium sevillanum Linsenmaier, 1968

Holopyga fastuosa Lucas, 1849

Holopyga jurinei sensu Linsenmaier 1959

Chrysis castillana du Buysson, 1894

Chrysis cerastes Abeille de Perrin, 1877

Chrysis insperata Chevrier, 1870

Chrysis crossi Rosa, sp. nov.

Stilbum westermanni Dahlbom, 1845

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Supplementary materials

Suppl. material 1: IBI - Hymenoptera 02 Chrysididae library - Specimen details

[doi](#)

Authors: Paolo Rosa, Thomas Wood, Sónia Ferreira

Data type: Specimen data records

Brief description: The file includes information about all records in BOLD for the IBI - Hymenoptera 02 library. It contains collecting and identification data. The data are as downloaded from BOLD in the tsv format, without further processing.

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Suppl. material 2: IBI - Hymenoptera 02 Chrysididae library - Specimen details

[doi](#)

Authors: Paolo Rosa, Thomas Wood, Sónia Ferreira

Data type: Specimen data records

Brief description: The file includes information about all records in BOLD for the IBI - Hymenoptera 02 library. It contains collecting and identification data. The data are as downloaded from BOLD in the DWC format, without further processing.

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Suppl. material 3: IBI - Hymenoptera 02 Chrysididae library - DNA sequences [doi](#)

Authors: Paolo Rosa, Thomas Wood, Sónia Ferreira

Data type: Genomic data, DNA sequences

Brief description: COI sequences in fasta format. Each sequence is identified by the BOLD ProcessID, species name, marker and GenBank accession number, separated by pipe. The data are as downloaded from BOLD.

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